

# Borrelia Biofilm: Is it real, and why is it important in chronic Lyme disease?

---

EVA SAPI PH.D.

UNIVERSITY OF NEW HAVEN

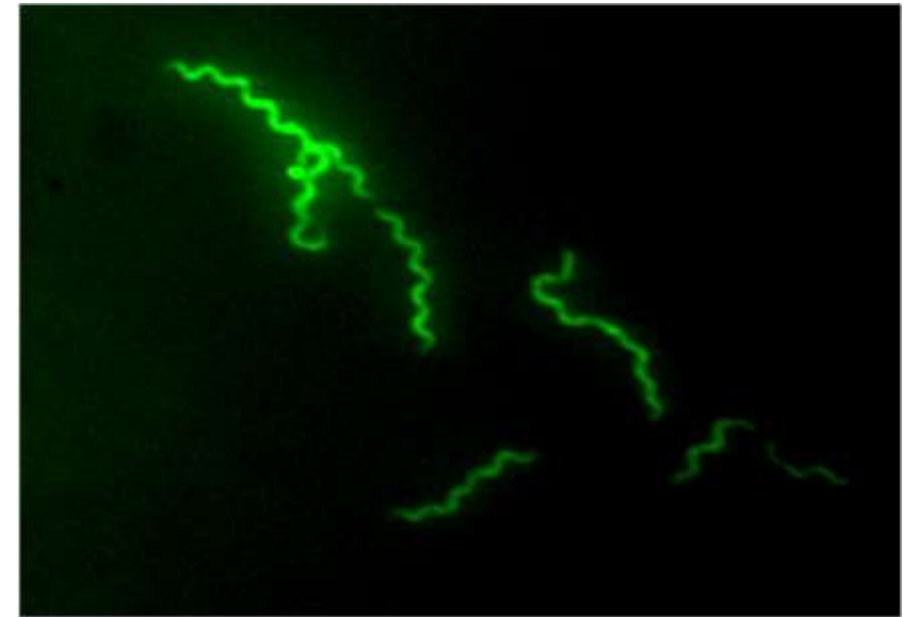


# *Borrelia burgdorferi* the spirochete that causes Lyme disease

---

In 1982, the etiologic agent of Lyme disease was discovered by **Dr. Willy Burgdorfer** who isolated spirochetes belonging to the genus *Borrelia* from the mid-guts of *Ixodes* ticks.

He showed that these spirochetes reacted with immune serum from patients that had been diagnosed with Lyme disease. Subsequently, the etiologic agent was given the name *Borrelia burgdorferi*.



*Borrelia burgdorferi* – University of New Haven

# How long did we know about Lyme disease?

---

John Josselyn, who visited New England in 1638 and again from 1663–1670, wrote

"there be infinite numbers of tikes hanging upon the bushes in summer time that will cleave to man's garments and creep into his breeches eating themselves in a short time into the very flesh of a man. I have seen the stockings of those that have gone through the woods covered with them"

The examination of preserved museum specimens has found *Borrelia* DNA in an infected *Ixodes ricinus* tick from Germany that dates back to 1884, and from an infected mouse from Cape Cod that died in 1894.

The 2010 autopsy of *Ötzi the Iceman*, a 5,300-year-old mummy, revealed the presence of the DNA sequence of *Borrelia* making him the earliest known human with Lyme disease.



---

So how can we eliminate *Borrelia*?

## *In vitro* studies for different species of *Borrelia* – Minimum bactericidal concentrations (microgram/ml)

---

Doxycycline:	0.25- <b>25.0</b>
Penicillin	0.15- <b>6.5</b>
Azitromycin:	0.015- <b>2.0</b>
Erythromycin	0.06->0.5
Clarithromycin	0.06-0.5
Telitromycin	0.002-0.03
Amoxicillin:	0.4- <b>8.00</b>
Ceftriaxone:	0.03-2.00*
Ciproflaxin	0.5- <b>16.0</b>
Tigecycline	0.05-0.19

◦Russel et al 1987, Agger et al 1992; Dever et al 1992, Levin et al 1993; Sicklinger et al 2003, Hunfeld et al 2004, 2005\*, Kim et al 2006, Yang et al 2009, Brorson et al 2009

# *In vitro* and *clinical* data – do they agree?

---

“*Survival of **Borrelia burgdorferi** in patients with Lyme borreliosis treated with antibiotics*”  
*Preac-Mursic V et al 1989*

“*In vitro* results have no proven correlation with antimicrobial clinical effectiveness *in vivo*”  
*Moody KD et al 1994*

“*Culture positive and PCR positive blood* after antibiotics therapy”  
*Oksi J et al 1999*

“*Clinically treatment failures occur in 5 to 10% of EM patients* (oral doxycycline or amoxicillin for 14 to 30 days)”  
*Smith RP et al 2002*

## But how about the *in vivo* studies?

---

Treatment with tetracycline, erythromycin or doxycycline in mice **failed to eradicate** acute *Borrelia* infection.

*Moody KD et al 1994*

Chloramphenicol and azithromycin **failed to eradicate** the organism - *Moody KD et al 1994*

In a dog model, results showed that antibiotic-treated dogs (doxycycline and amoxicillin, 30 days) continued to have **persistent *Borrelia*-specific DNA** in their tissue albeit at lower levels than observed in untreated animals.

*Straubinger RK et al 1997*

## But how about Ceftriaxone (Rocephin)?

---

Bockenstedt LK, Mao J, Hodzic E et al. Detection of **attenuated, noninfectious spirochetes** in *Borrelia burgdorferi*-infected mice after antibiotic treatment. J Infect Dis 2002; 186: 1430–7.

Hodzic E, Feng S, Holden K et al. **Persistence of *Borrelia burgdorferi*** following antibiotic treatment in mice. Antimicrob Agents Chemother 2008; 52: 1728–36.

SUMMARY: **A low numbers of noncultivable spirochetes**, detected by PCR following antibiotic treatment which can be **acquired by ticks, transmitted by ticks**, survive the molts between larvae to nymphs to adults, infect mice, and **form of morphologically identifiable spirochetes**



# The different forms of Borrelia

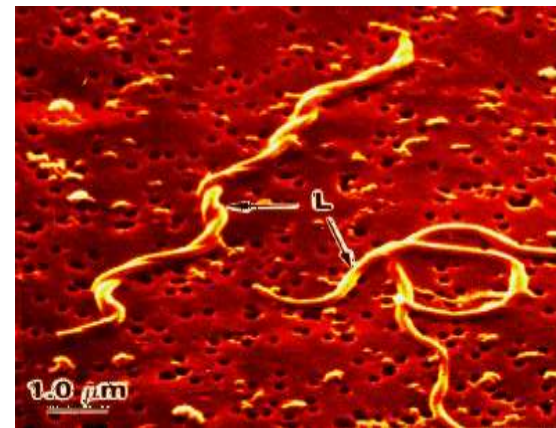
*Borrelia burgdorferi* can convert between cyst, non-motile and normal motile spirochete forms.

The cystic forms are **resistant** to most antibiotic treatments and difficult to detect in the body.

- <http://www.lymeinfo.net/medical/LDAdverseConditions.pdf>



*B. burgdorferi* after exposure to penicillin concentration of 0.125 mg/l. Coiled up spirochete forming a spherical structure (spheroplast).  
Schaller M; Neubert U. 1994



# The most recognized forms of *Borrelia burgdorferi*

- Spirochetes

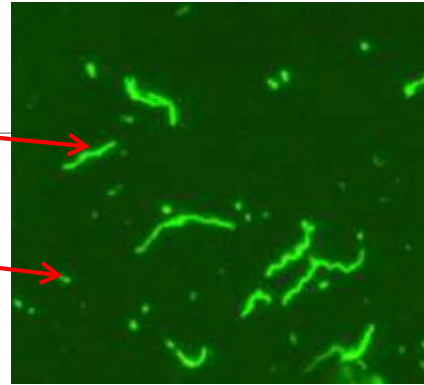


Photo by Namrata Pabbati MS

- Round bodies (cysts, granules)



Photo by Alan MacDonald MD, 2006

- L-forms

- Biofilm

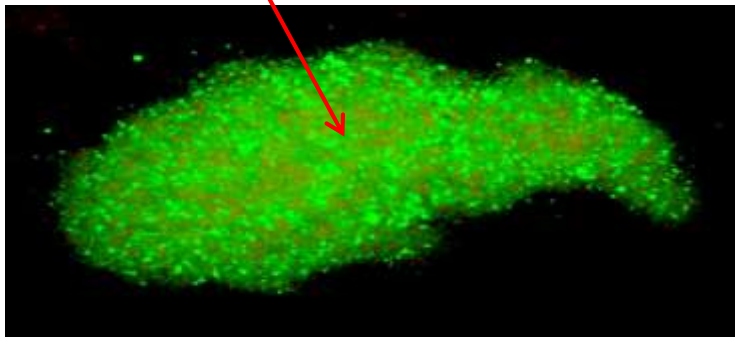


Photo by David Luecke MS

## Round bodies *in vivo*

---

### Neocortical Borreliosis and Alzheimer's Disease Demonstration of a Spirochetal Cyst Form

*MacDonald A 1988*

- An unexpected observation was the identification of cystic forms of the *Borrelia* spirochete by dark field microscopy cultured hippocampus

### Conversion of *Borrelia garinii* Cystic Forms to Motile Spirochetes *In Vivo*.

*Gruntar I et al 2001*

- *Borrelia garinii* cystic forms maintain their capability to reconvert into normal spirochetes not only *in vitro* but also *in vivo* and can therefore be considered infective, at least in BALB/c mice.

## Agents for the cystic forms (RB)

---

*In Vitro* Study of the Susceptibility of Mobile and Cystic Forms of *Borrelia burgdorferi* to **Metronidazole** Brorson O et al 1999

“*B. burgdorferi* has the ability to make cystic forms both *in vivo* and *in vitro*, e.g. when exposed to antibiotics commonly used for treating Lyme borreliosis. This phenomenon, combined with the ability of the **cysts to reconvert to normal mobile spirochetes** may explain a reactivation of the disease— and not a “post Lyme syndrome” as postulated by other researchers.”

## Additional Brorson O et al *in vitro* studies for the antibiotic sensitivity of the cystic (round bodies) form

---

**2001** Susceptibility of motile and cystic forms of *Borrelia burgdorferi* to **ranitidine bismuth citrate**. Int Microbiol, 4(4):209-15.

**2002** An *in vitro* study of the susceptibility of mobile and cystic forms of *Borrelia burgdorferi* to **hydroxychloroquine** Int Microbiol, 5(1):25-31.

**2004** An *in vitro* study of the susceptibility of mobile and cystic forms of *Borrelia burgdorferi* to **tinidazole**. Int Microbiol, 7(2):139-40.

**2009** Destruction of spirochete *Borrelia burgdorferi* round-bodies by **tigecycline** PNAS 106(44):18656-61.

## Ineffectiveness of Tigecycline against Persistent *Borrelia burgdorferi* *in vivo*

---

**Non-cultivable** *Borrelia burgdorferi* could be isolated from mice treated with ceftriaxone and tigecycline.

Mice remained **infected with non-dividing, but infectious spirochetes**, particularly when antibiotic treatment was commenced during the chronic stage of infection

*Barthold SW et al 2010*



Embers ME et al:

Persistence of *Borrelia burgdorferi* in Rhesus Macaques following Antibiotic Treatment of Disseminated Infection. PLoS ONE 2012

---

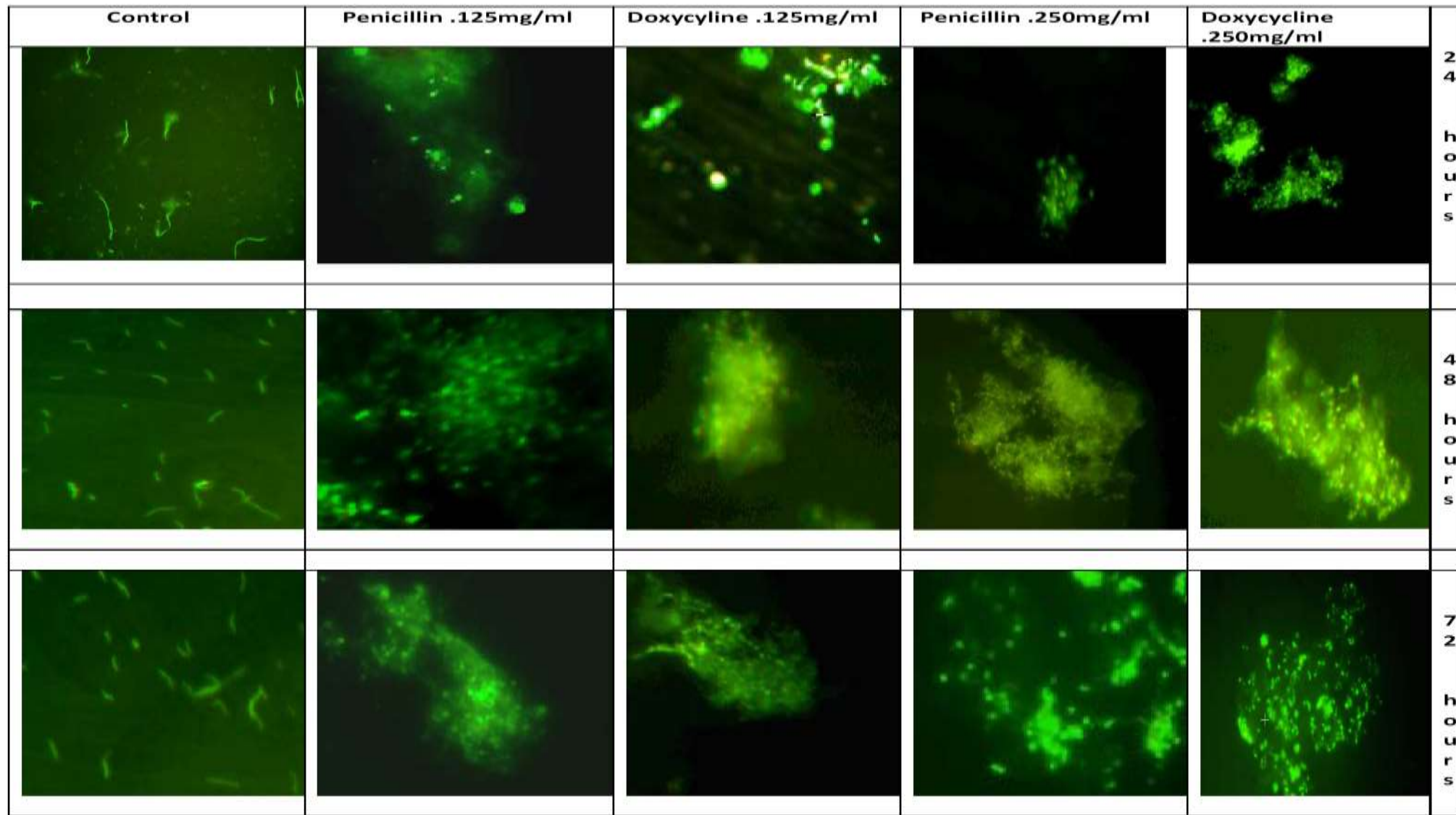
- found that **Borrelia persist** after 90 days in monkeys treated for chronic Lyme disease.
- the antibody tests used to diagnose Lyme disease **fail to detect disease in late Lyme disease** at least 50% of the time.

# So what can we do now ????

*What other escape route Borrelia could have???*



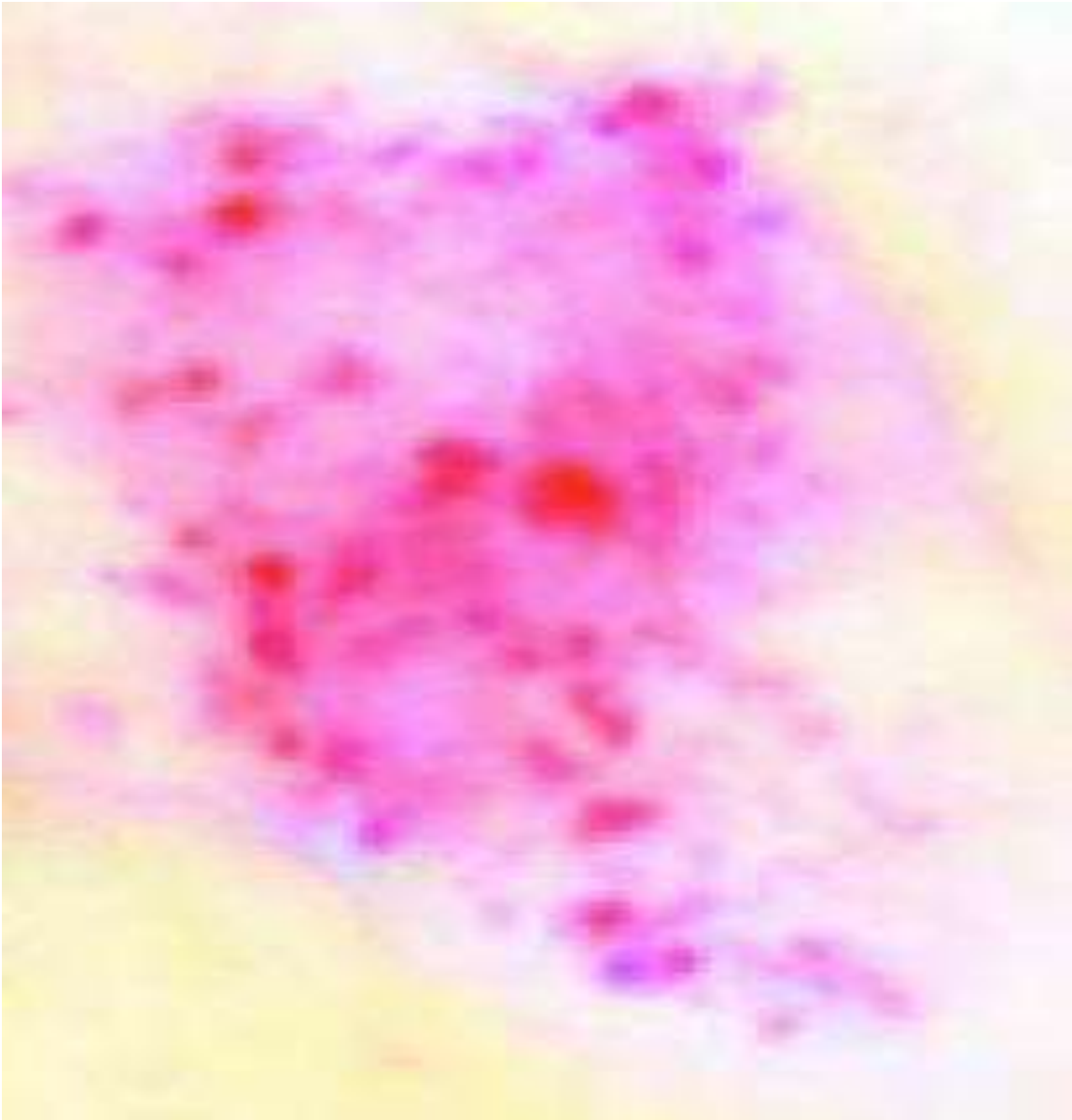


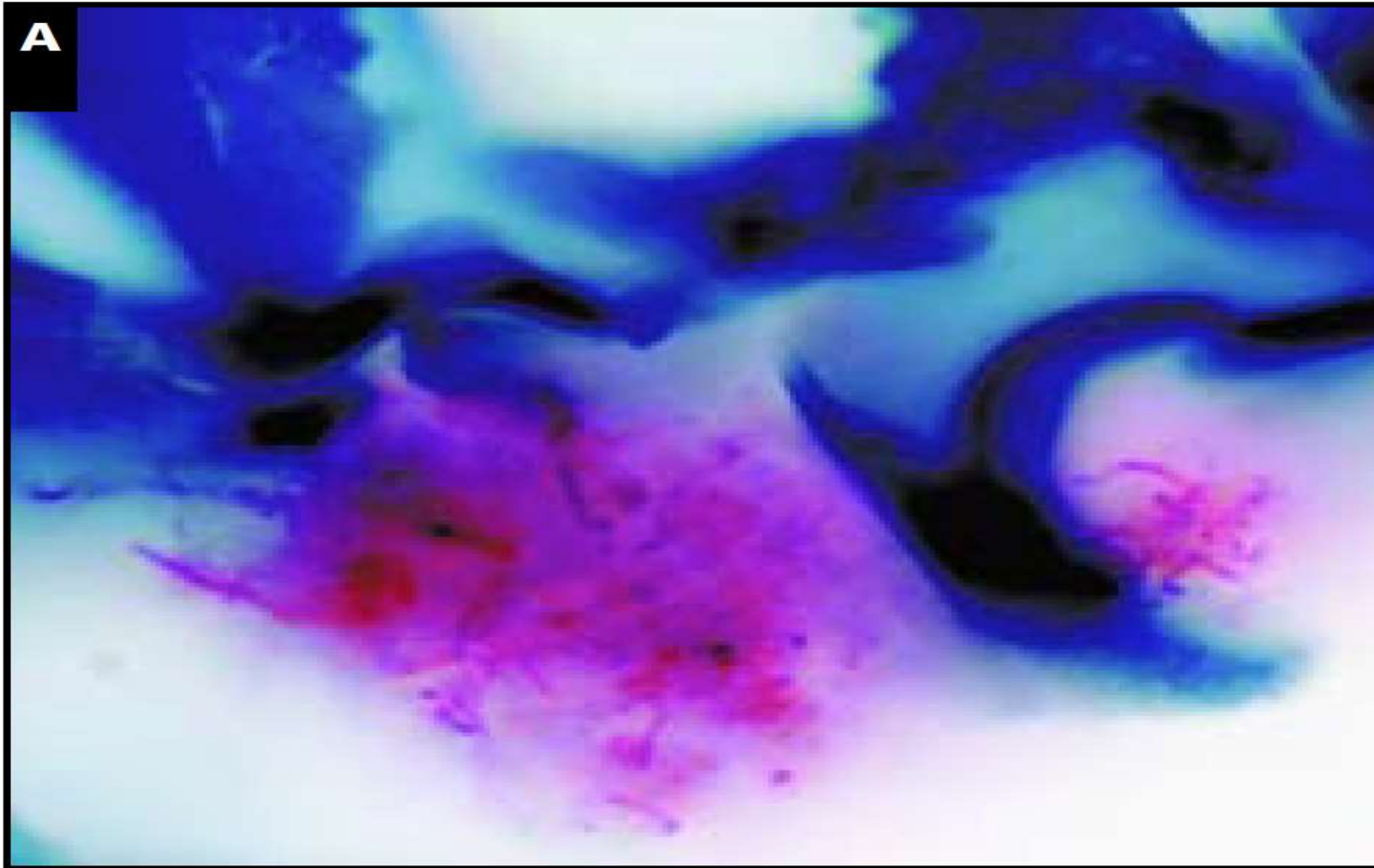


Rao P, Azano, D & Sapi E, unpublished data 2008

K. Eisendle et al. AJCP 2007,127:213-222 Acrodermatitis Chronica Atrophicans Immunohistochemistry

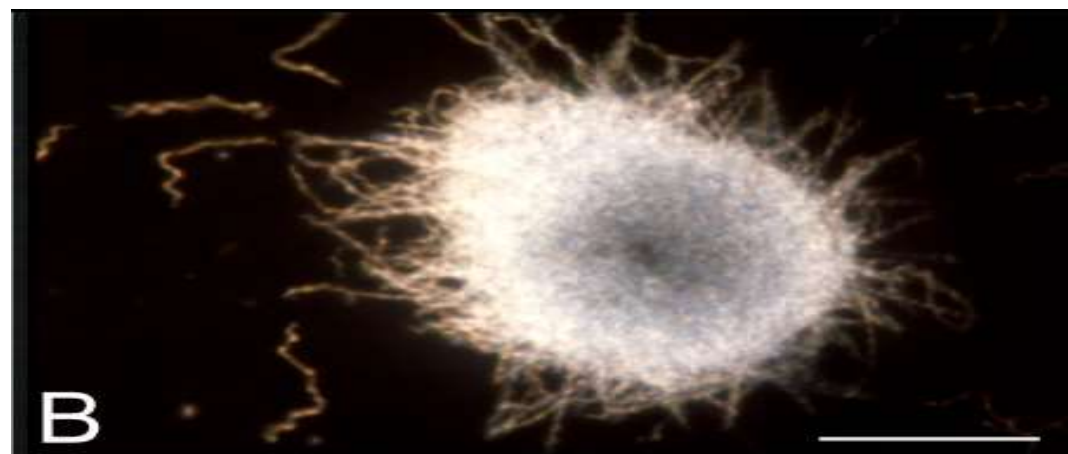
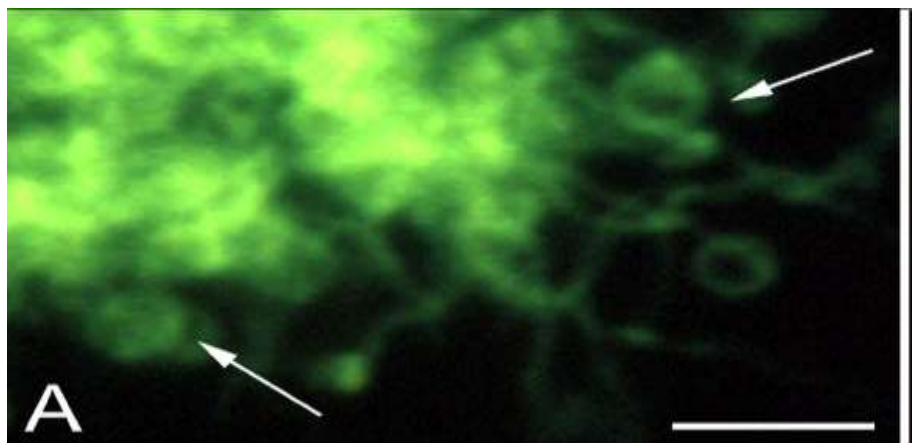
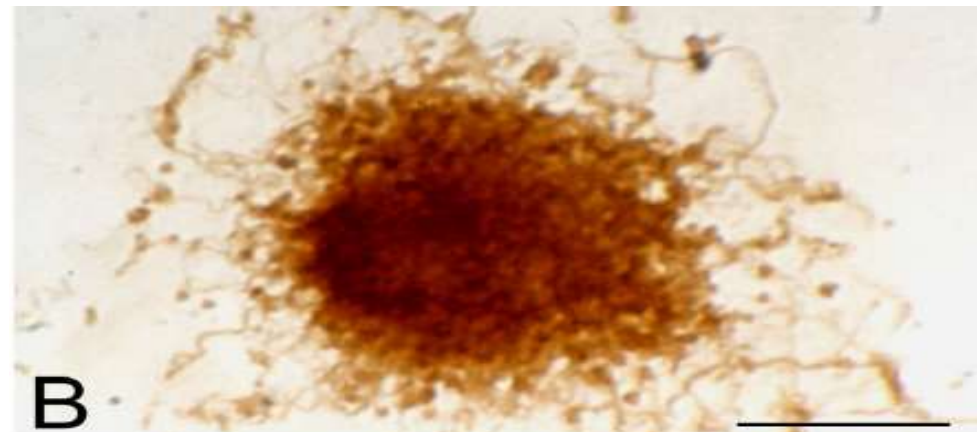
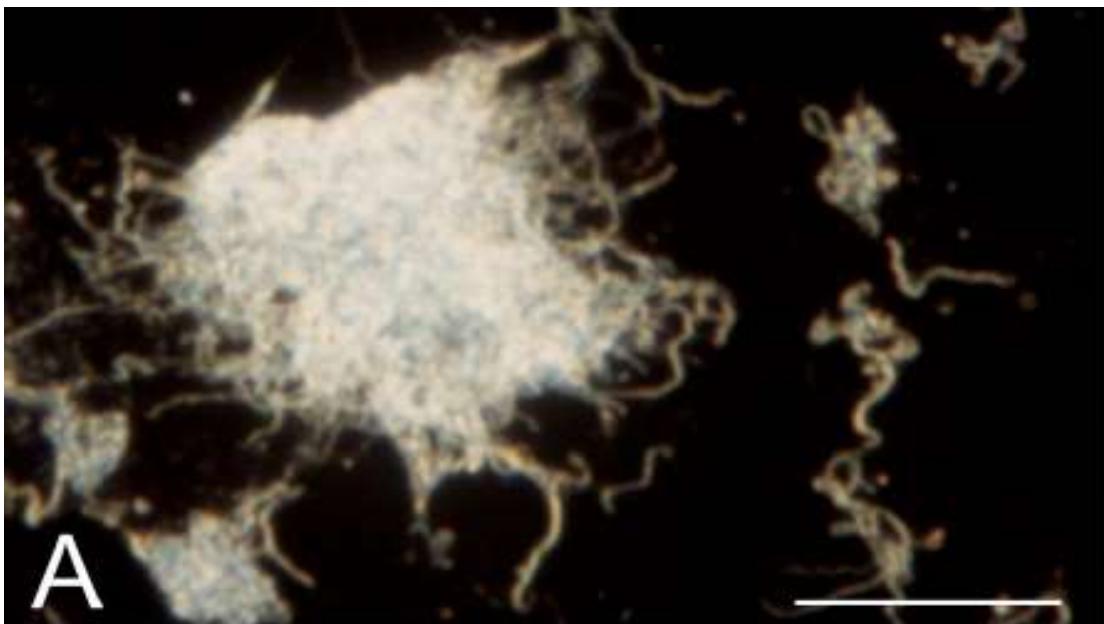
“Granular forms of *Borrelia burgdorferi* in a “colony” with a “reddish veil”





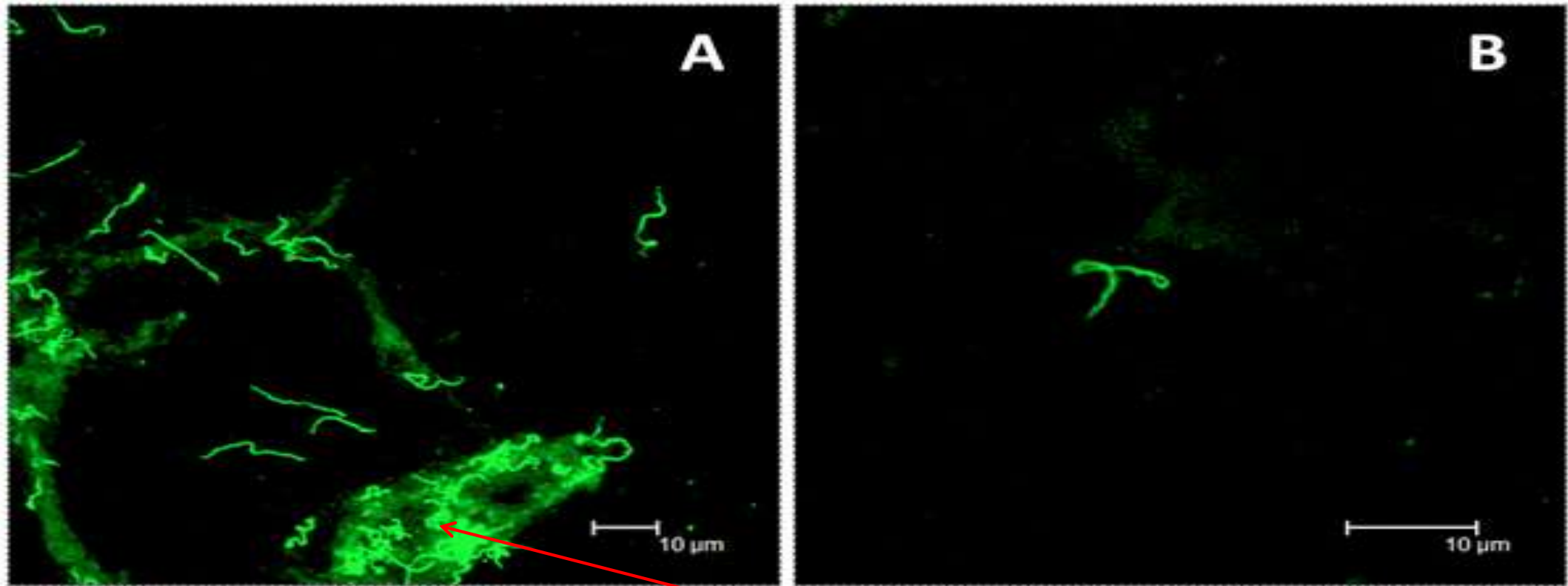
*Borrelial lymphocytoma with medusa-like colony of Borrelia*

*Borrelia burgdorferi* “colonies”



Miklossy J et al 2008

## The Rhesus Macaques' study: Embers ME 2012

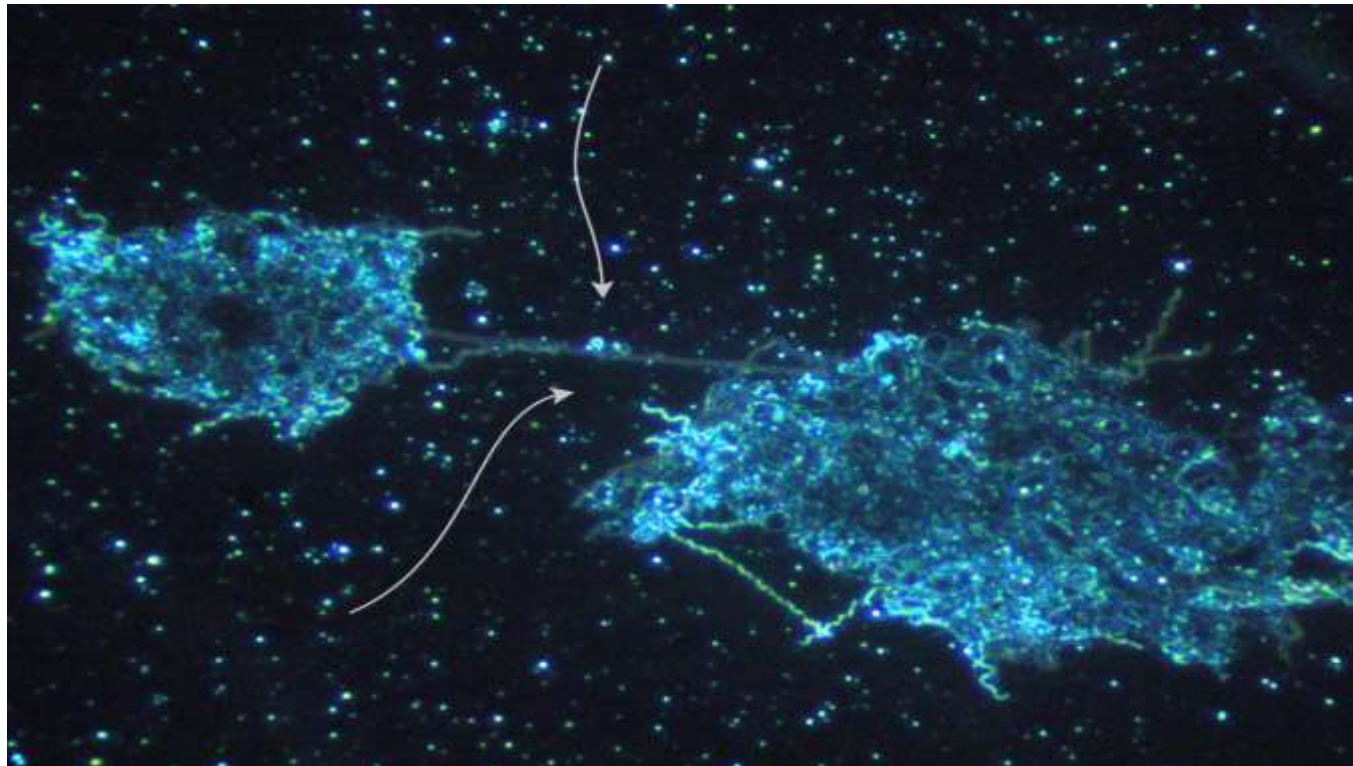


Fluorescent staining of *Borrelia burgdorferi* spirochetes found in xenodiagnostic tick midgut culture (A) or tick midgut preparation (B) from treated animals

*Borrelia* colonies or aggregates?

# *Borrelia burgdorferi* “Photo 51”

---



MacDonald A, & Sapi E: Biofilms of *Borrelia burgdorferi* in chronic cutaneous borreliosis  
AJCP 2008 June

---

- We propose the hypothesis that that *Borrelia burgdorferi* can form biofilm structures in lymphocytomas and acrodermatitis chronica atrophicans.
- Our close examination of these pictures revealed striking similarity to previously published biofilm pictures and our preliminary findings on specific biofilm-like colony formation of *Borrelia burgdorferi* when cultured in the presence of human plasma

# What is Biofilm?

---

**collections of microorganisms** (bacteria, yeasts or protozoa) that form on a hard surface (exception floating biofilms)

examples: plaque that forms on teeth and the slime that forms on surfaces in watery areas (shower)

**surrounded by slimy secretions**: mucoid polysaccharide structure which attaches the community to a surface

estimated that over 90% of bacteria live in biofilm (late Costerton WJ)



# Microorganisms found in medical devices

---

*Staphylococcus*

*Streptococcus*

*Enterococcus*

*E. coli*

*Klebsiella*

*Pseudomonas*

Bacteria may originate from the skin of the patient, or a healthcare worker and tap water

# Common biofilms

---

Dental plaque

Bacterial endocarditis

Urinary tract infections

Cystic fibrosis

Staphylococcus osteomyelitis

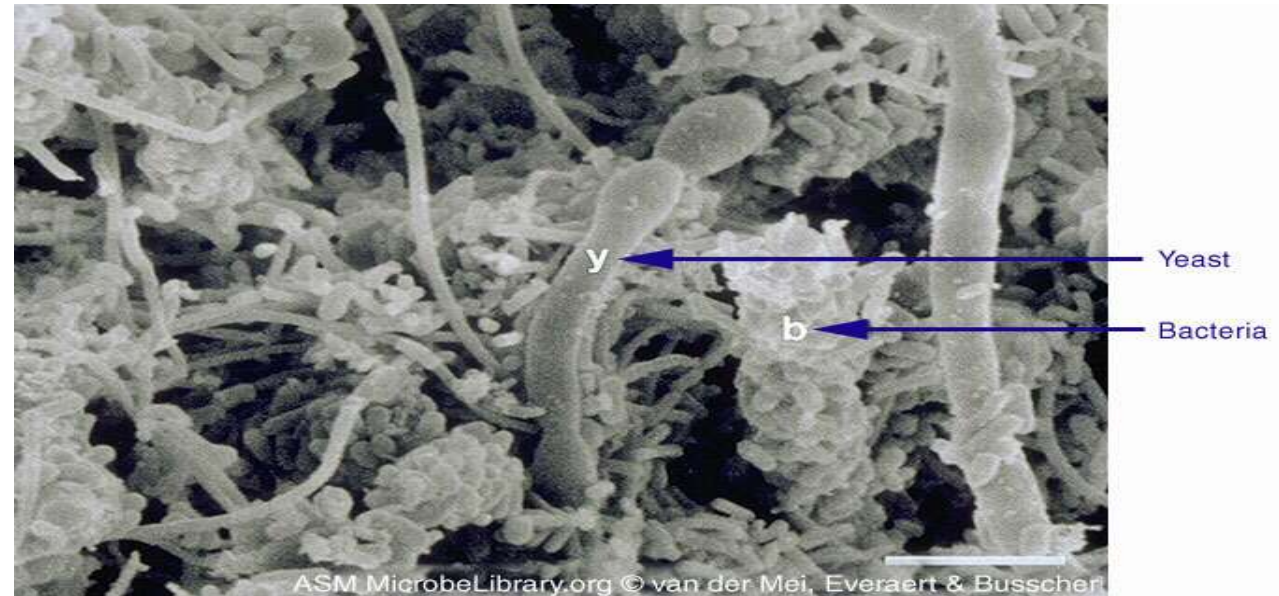
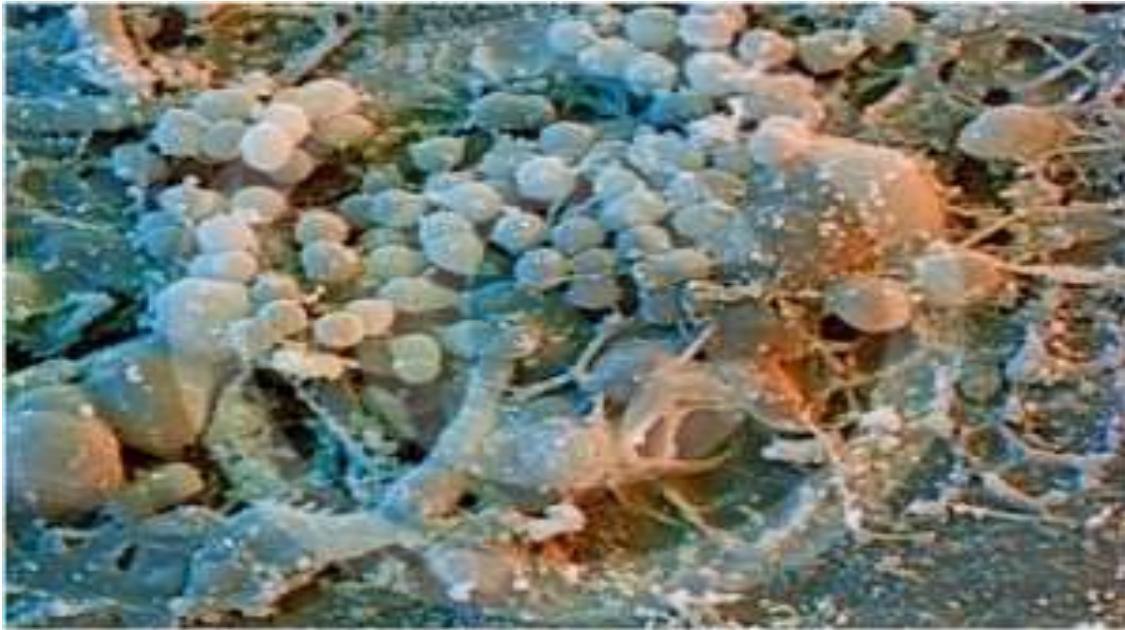
Middle ear infection

Chronic prostatitis

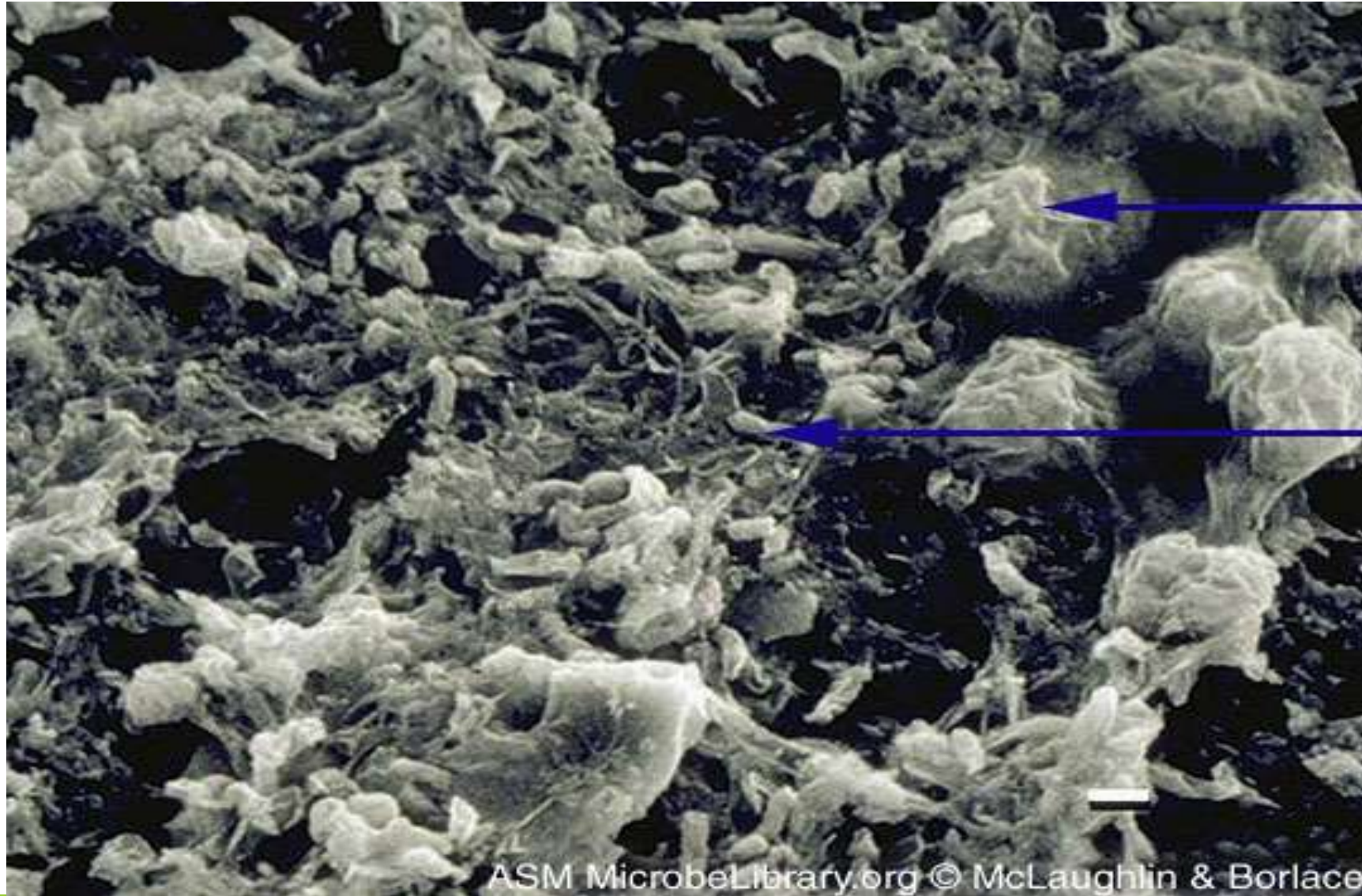
Infectious kidney stones

# Dental Plaque: Complex community

---



# Contact lens



Dried *Acanthamoeba*  
cysts

Rod-shaped  
bacteria

# Biofilm is like a city

---

Careful selection of location

Limit settlement of too many organisms

Division of labor (planktonic and sessile cells)

Storage of energy (exopolysaccharides)

Transfer of information (genetic transfer)

Intercellular communication

Emigration when population gets too large for resources

# What makes a bacterial colony to a true biofilm?

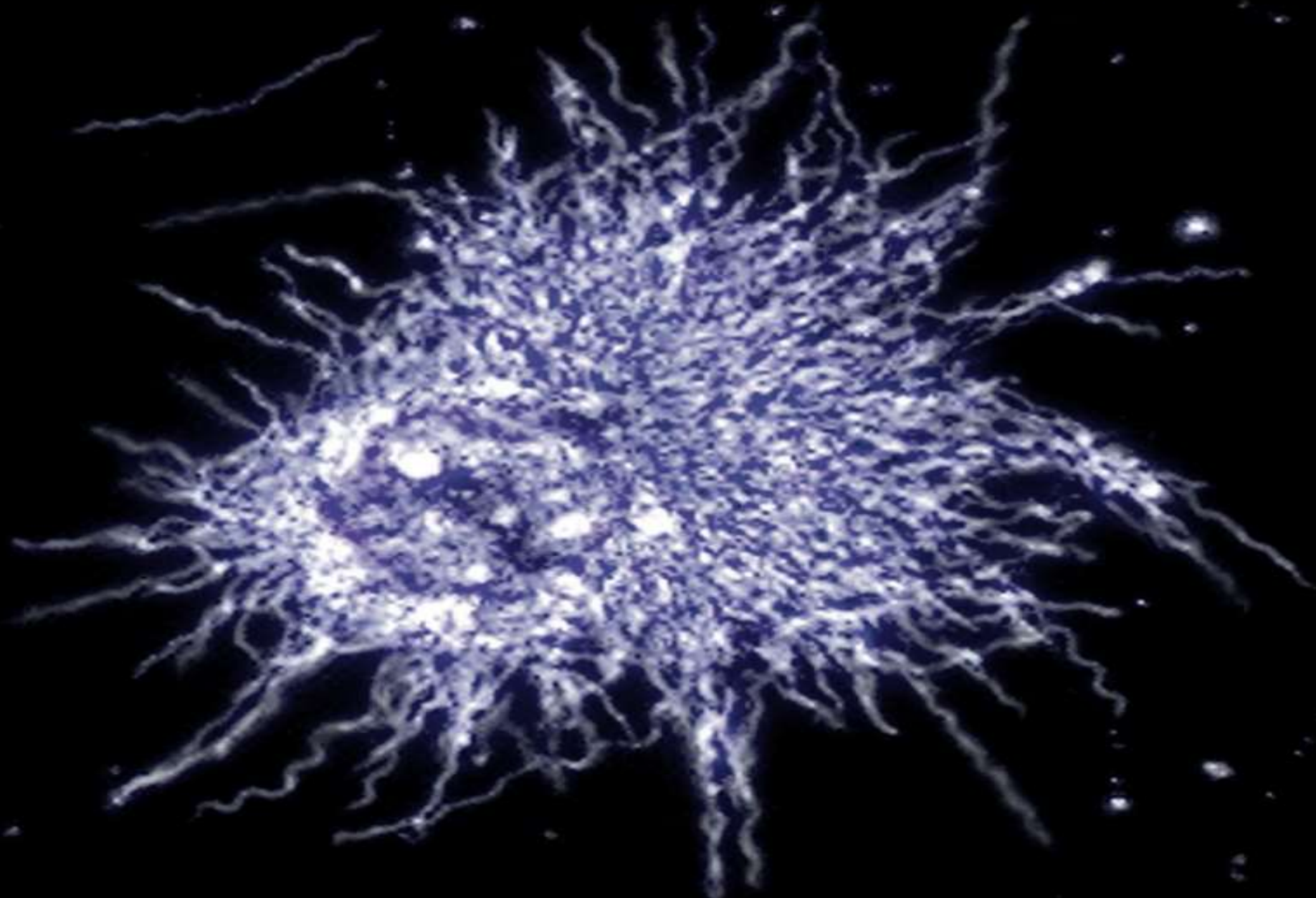
---

Loss of motility?

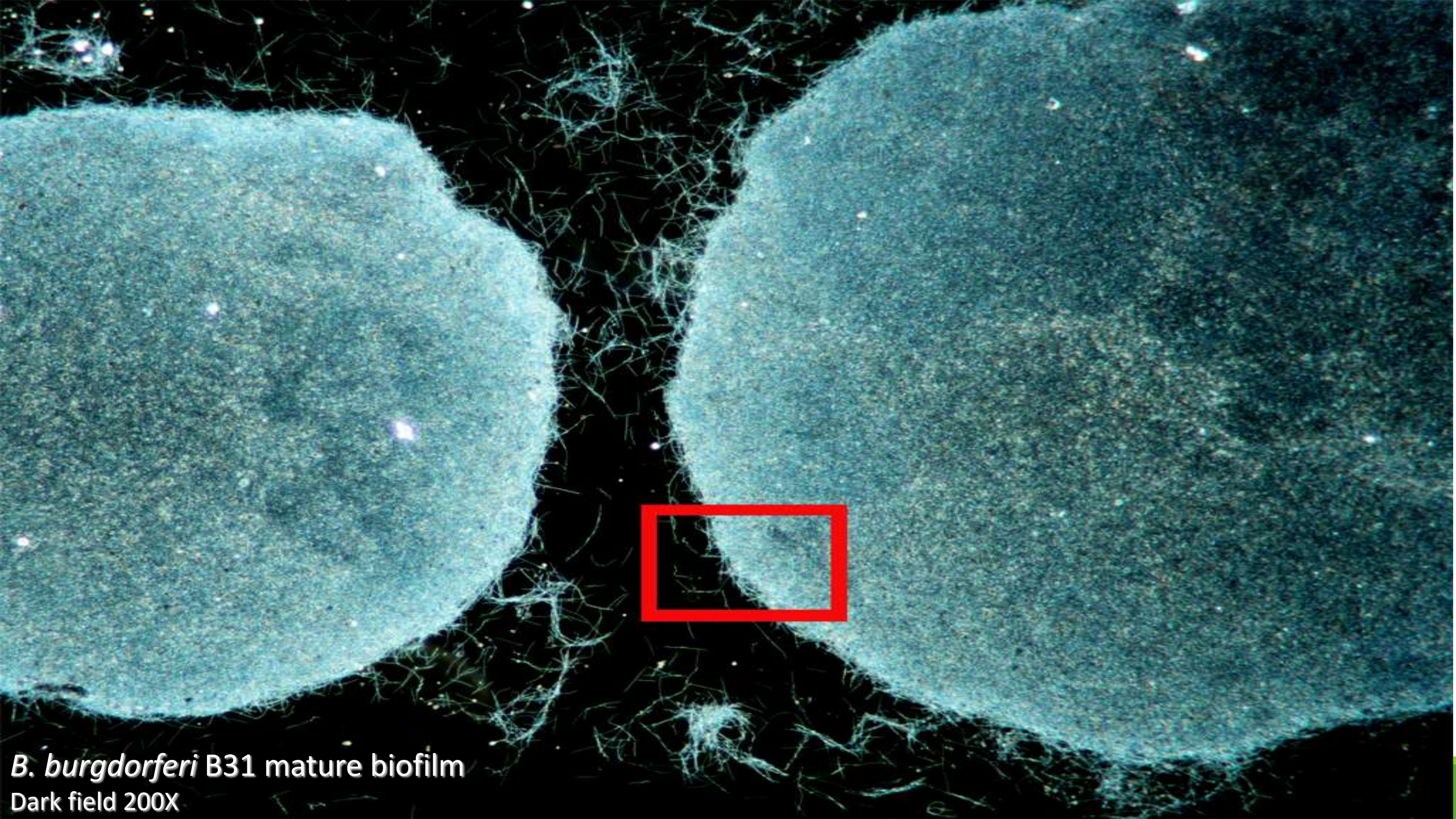
Internal morphological rearrangement

A colony embedded in a matrix of extracellular polymeric substance (EPS) separated by a network of open channels

Communication network- quorum sensing

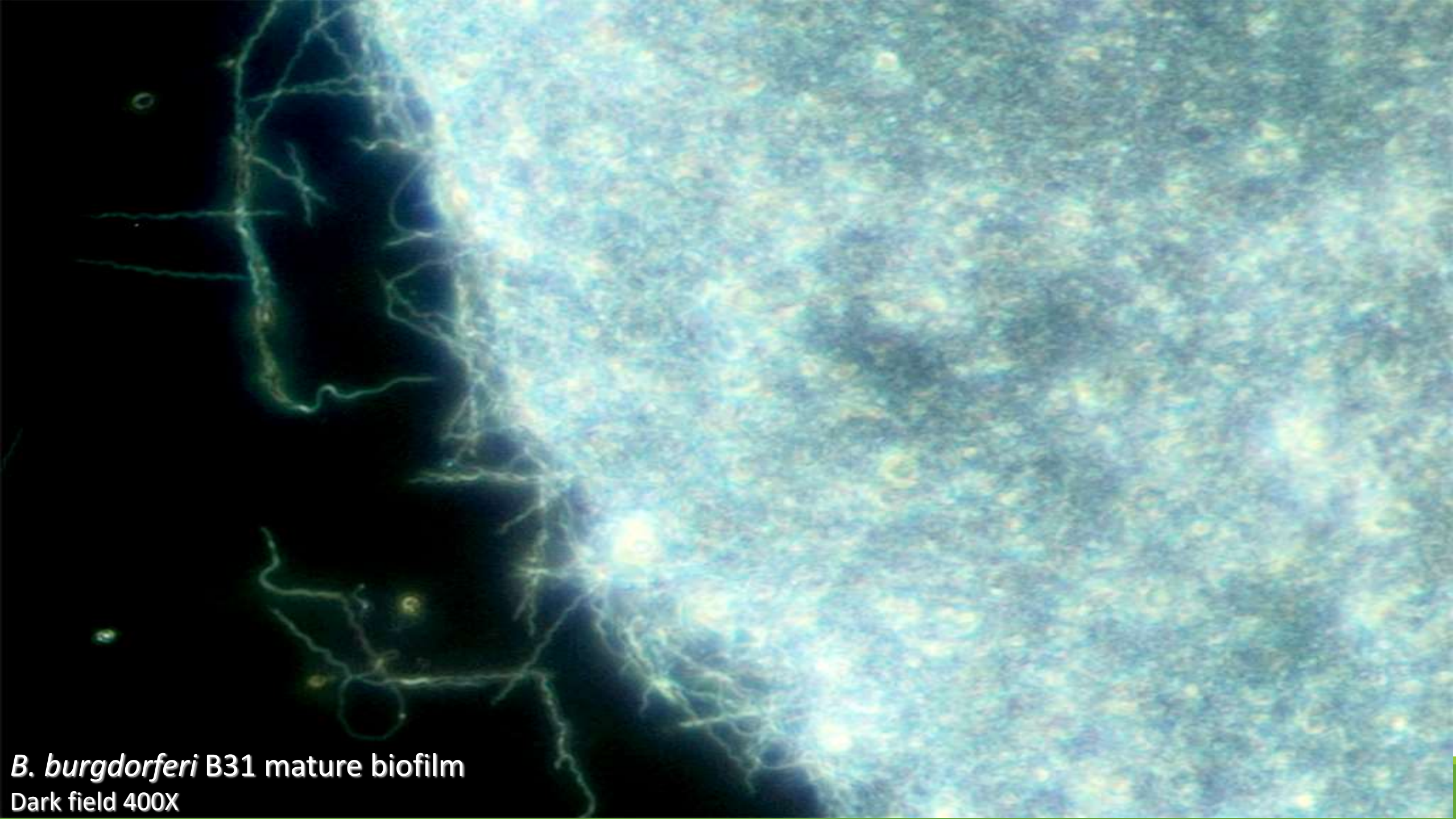


*B. burgdorferi* early development of biofilm-like structure  
dark field 40X



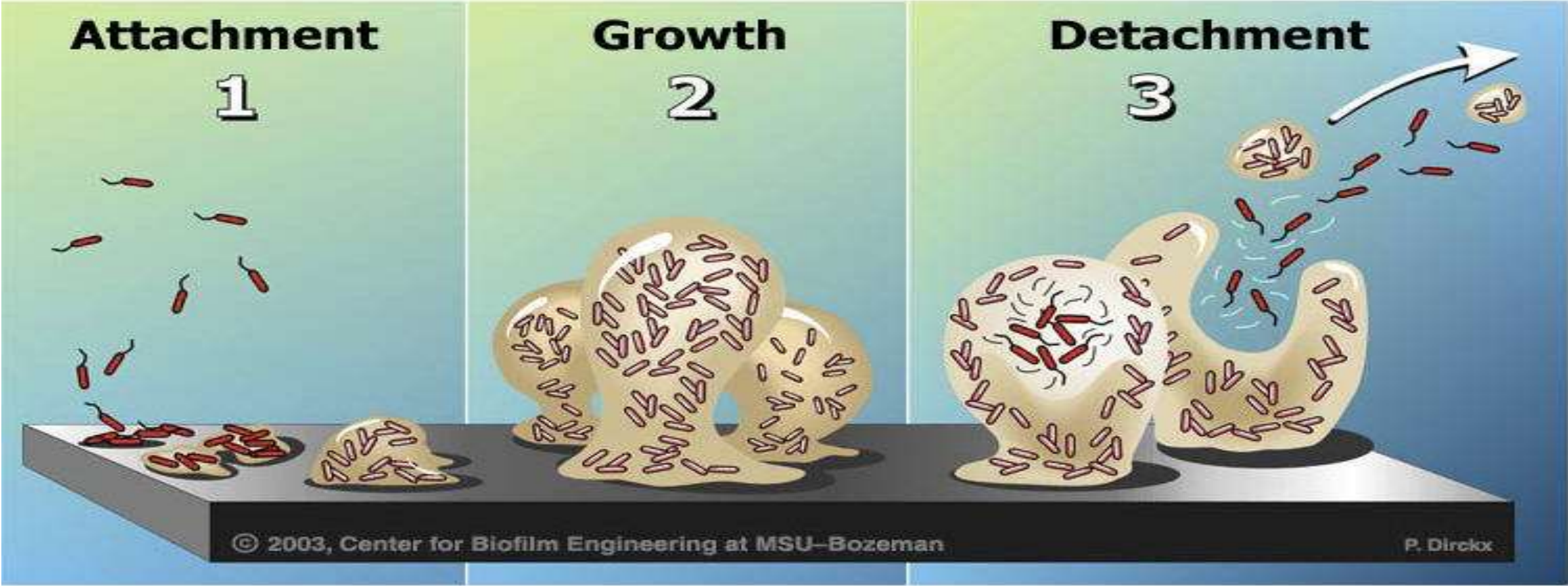
*B. burgdorferi* B31 mature biofilm  
Dark field 200X





*B. burgdorferi* B31 mature biofilm  
Dark field 400X

# Stages of biofilm development

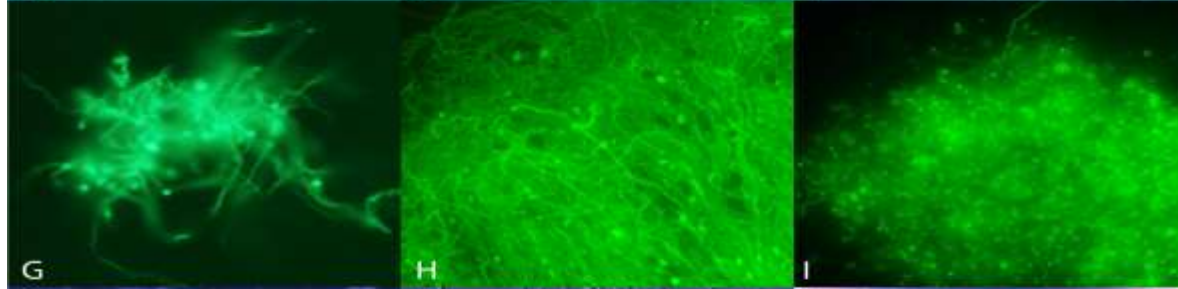




Dark-field  
microscope



DIC



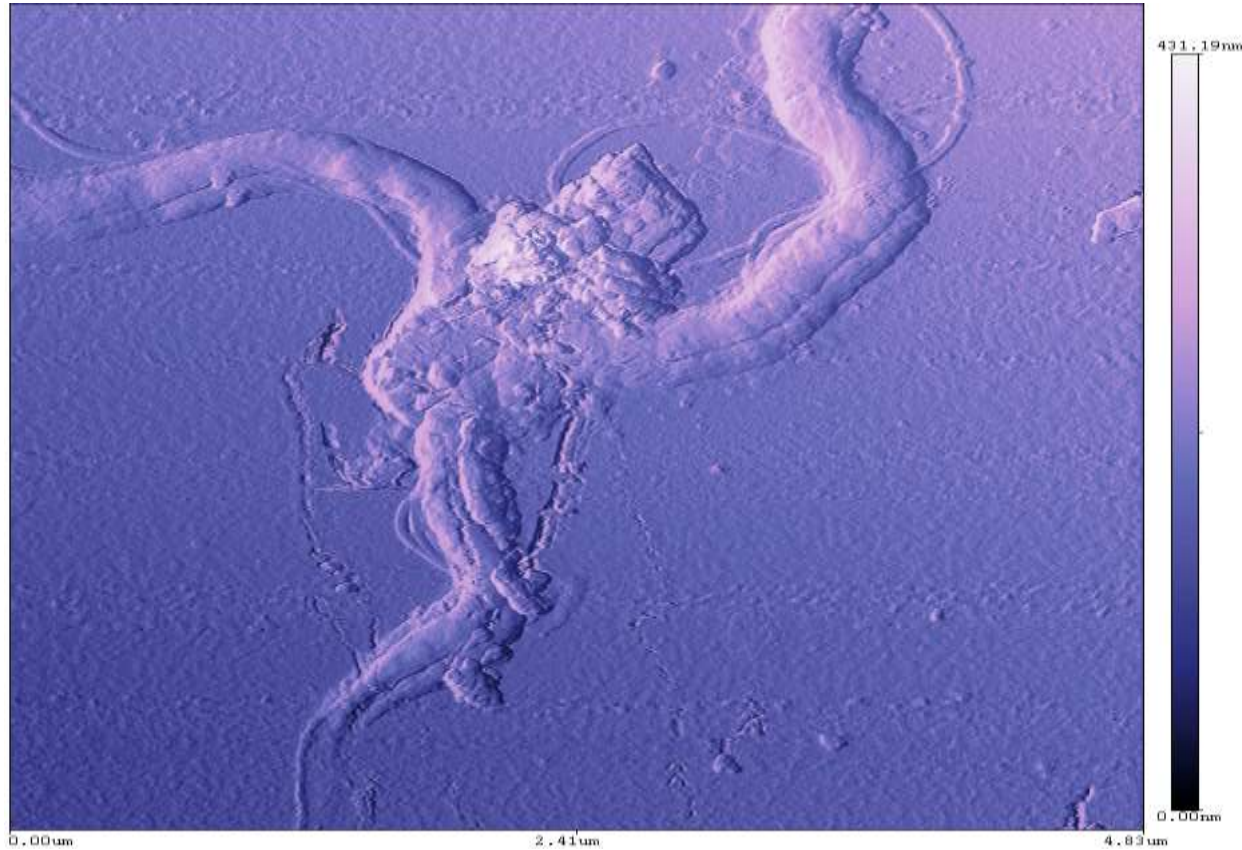
Fluorescent  
microscope



Atomic Force  
Microscope

*Borrelia burgdorferi* biofilm Sapi E et al: PLoS One 2012

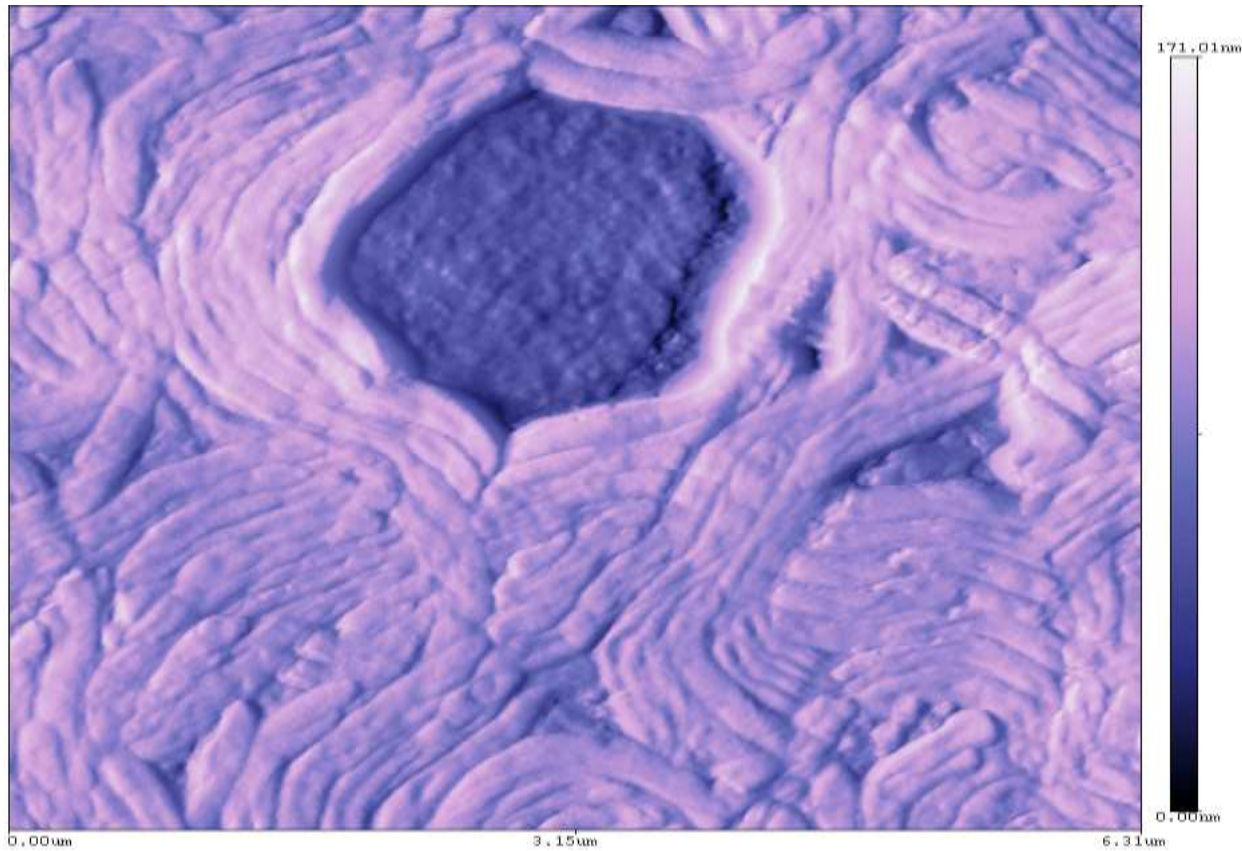
# Atomic force microscopy images of live *Borrelia* colony growing on agarose – early development



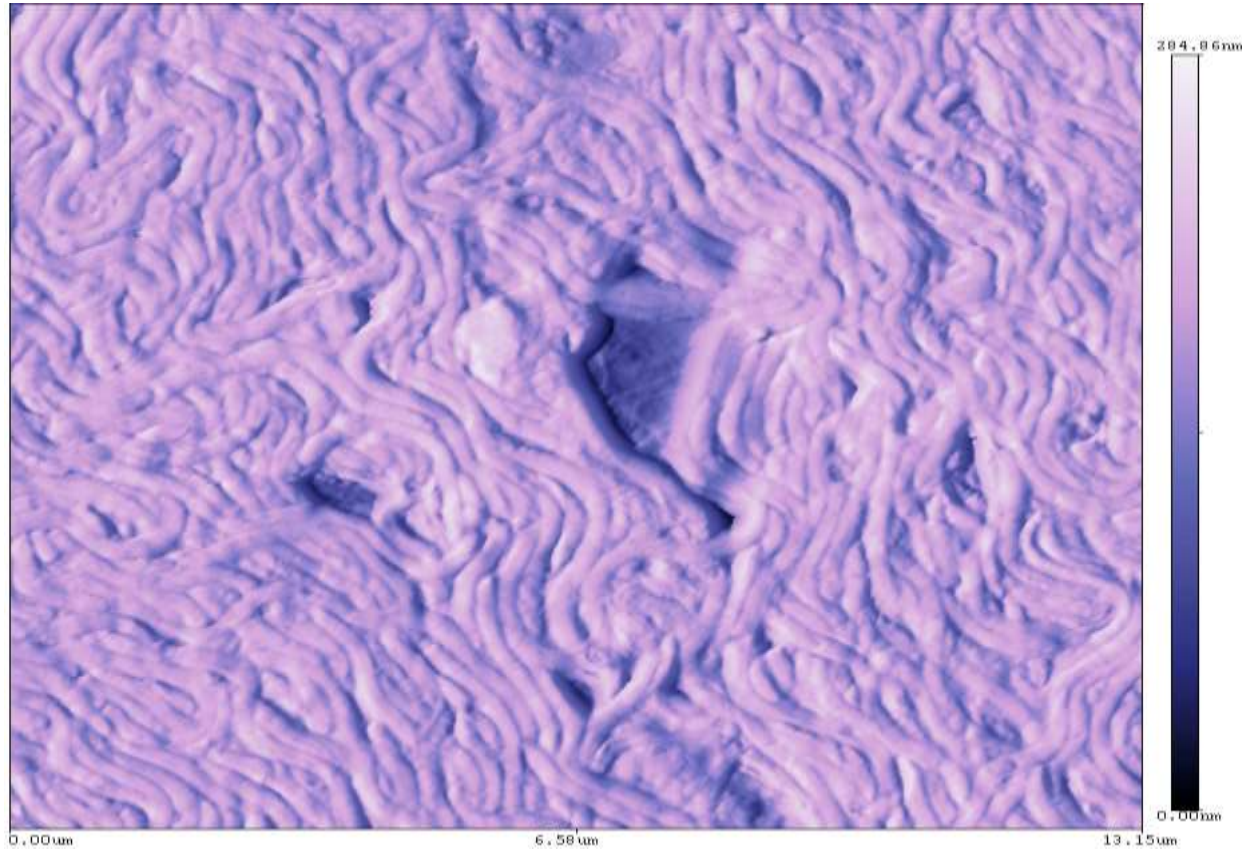
# Atomic force microscopy images of live *Borrelia* colony growing on agarose – early development



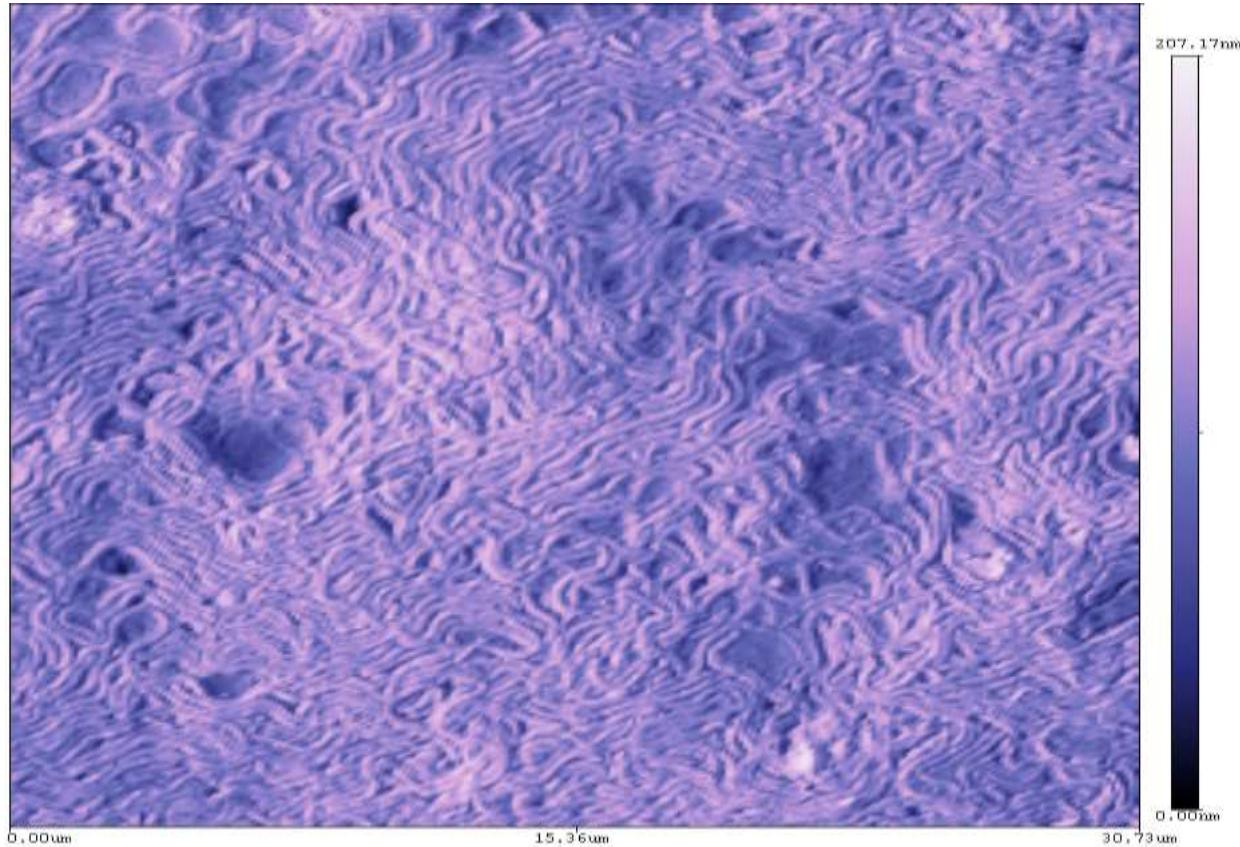
# Atomic force microscopy images of live *Borrelia* colony growing on agarose – mid-phase development



Atomic force microscopy images of live *Borrelia* colony growing on agarose – midphase development – 2 days later

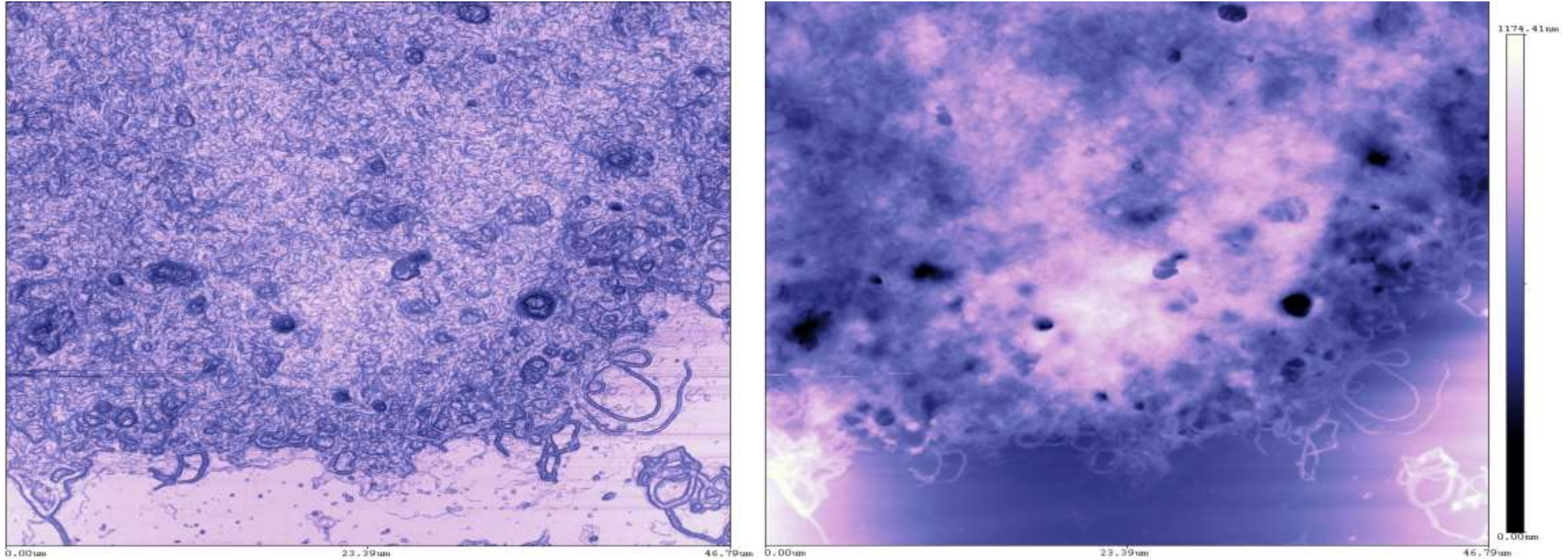


# Atomic force microscopy images of live *Borrelia* colony growing on agarose –late phase

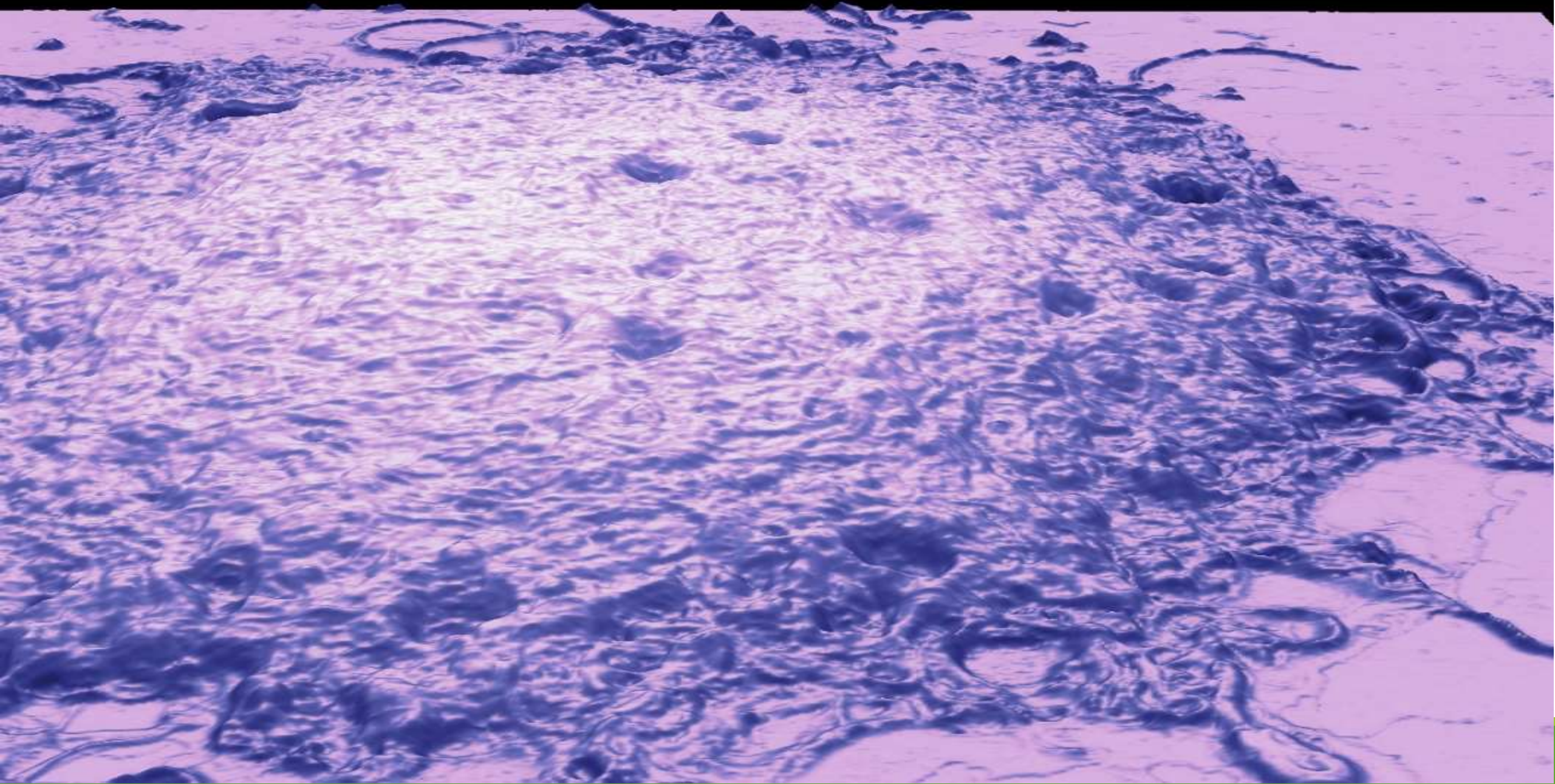


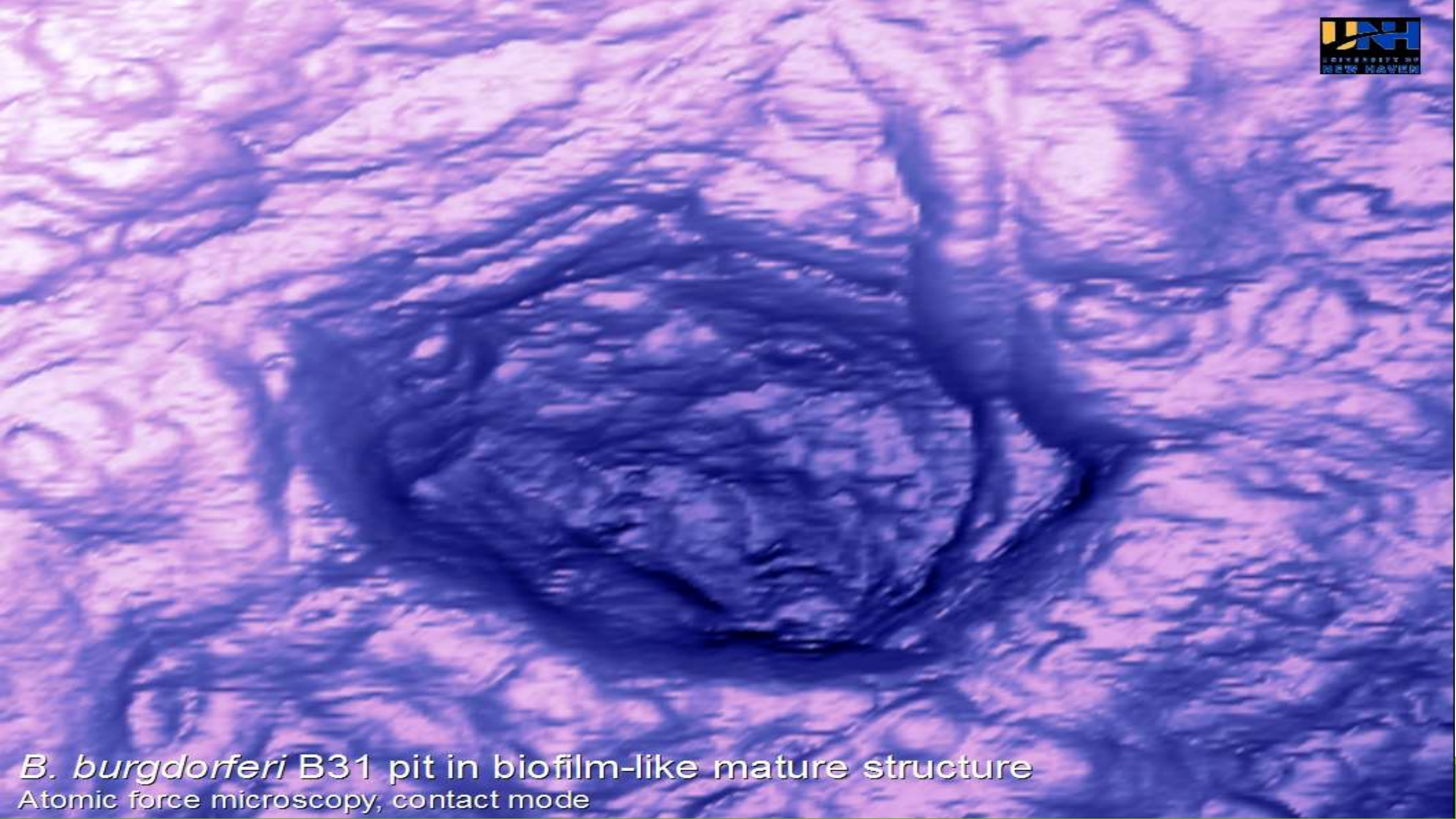


# Atomic force microscopy images of live *Borrelia* colony growing on agarose –late phase



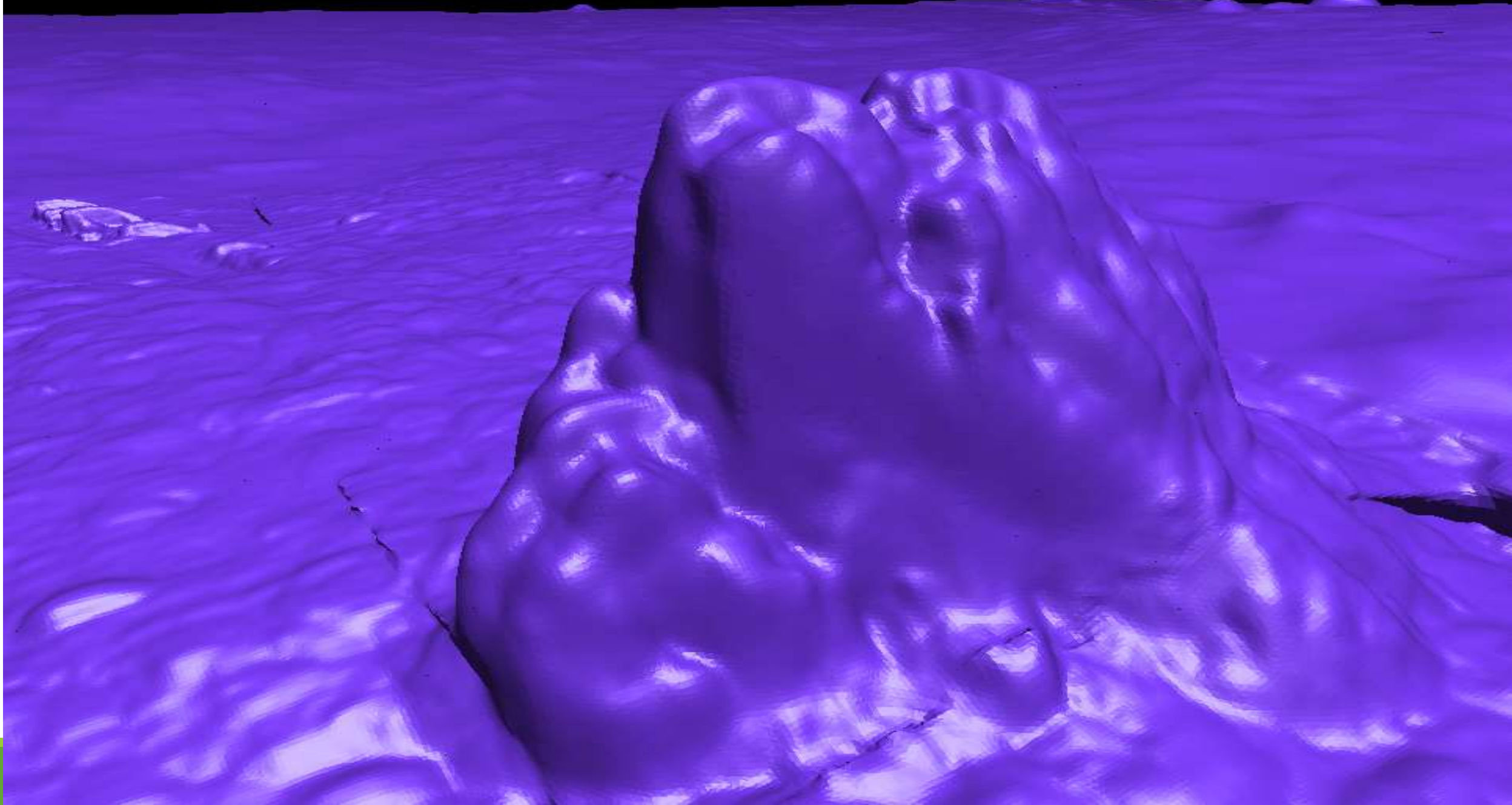
*B. burgdorferi* B31 biofilm-like mature structure  
Atomic force microscopy, contact mode



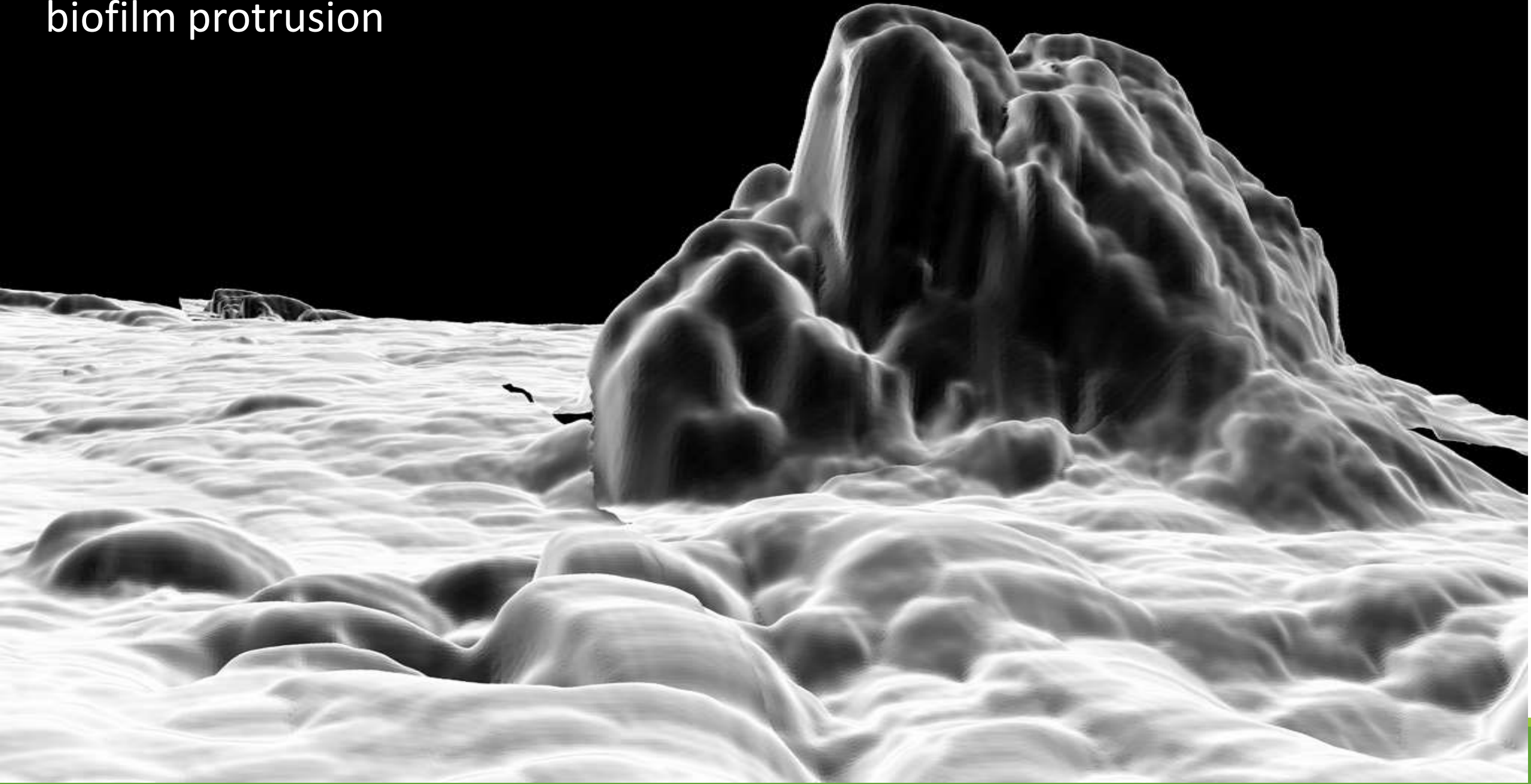


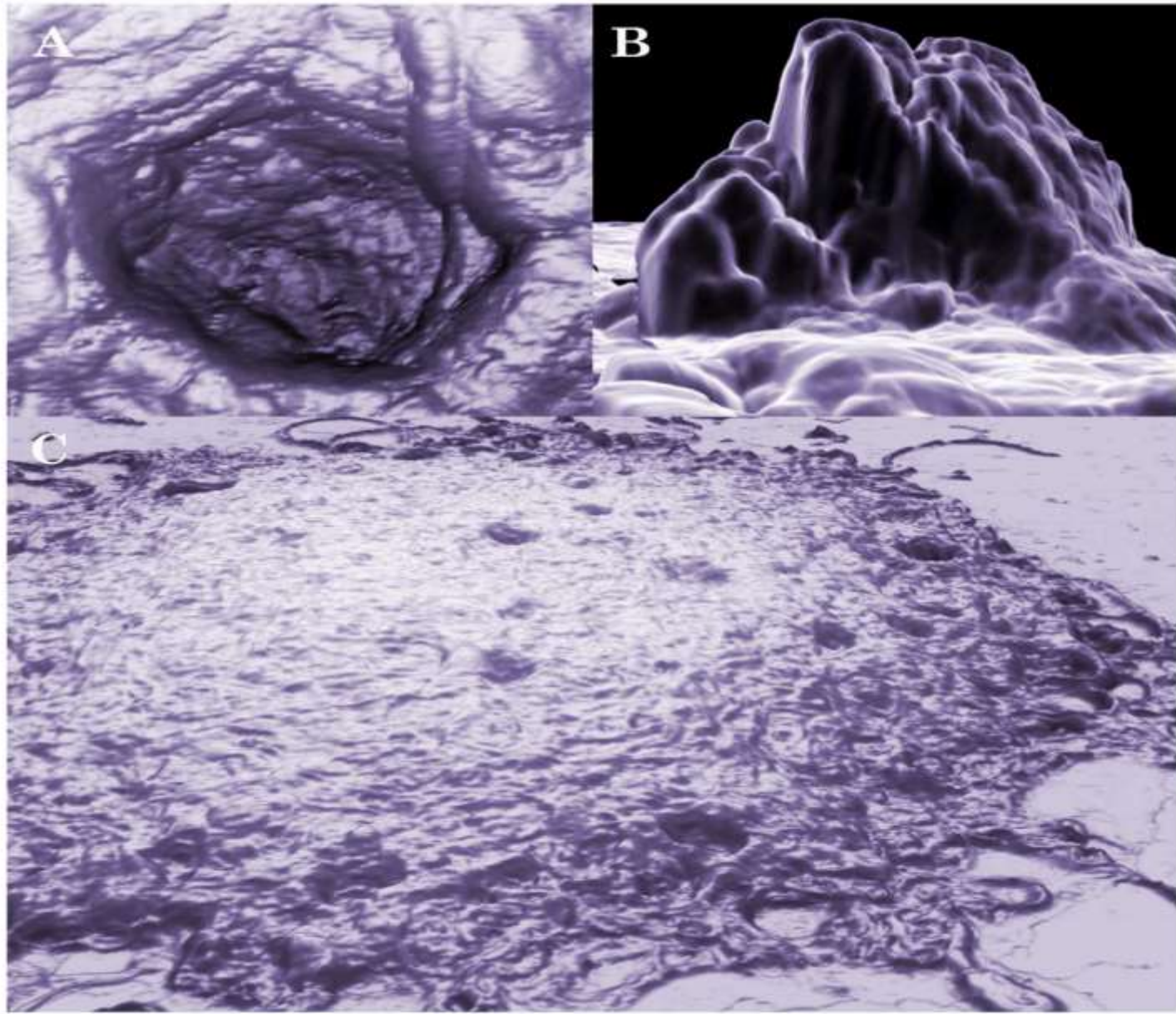
*B. burgdorferi* B31 pit in biofilm-like mature structure  
Atomic force microscopy, contact mode

*B. burgdorferi* biofilm protrusion



*B. burgdorferi*  
biofilm protrusion





# Extracellular Polymeric Substances

The EPS matrix: The “house of biofilm cells”

---

Composed of mucopolysaccharides (slime), proteins (enzymes) glycoproteins, glycolipids, extracellular DNA

Some of these polysaccharides are polyanionic (like uronic acid) – can bind to **calcium and magnesium**

Costerton JW and Irvin RT 1984, Flemming HC 2007

# Spicer Meyer aldehyde fuchsin – alcian blue stain sequence for mucopolysaccharides

---

## Aldehyde fuchsin:

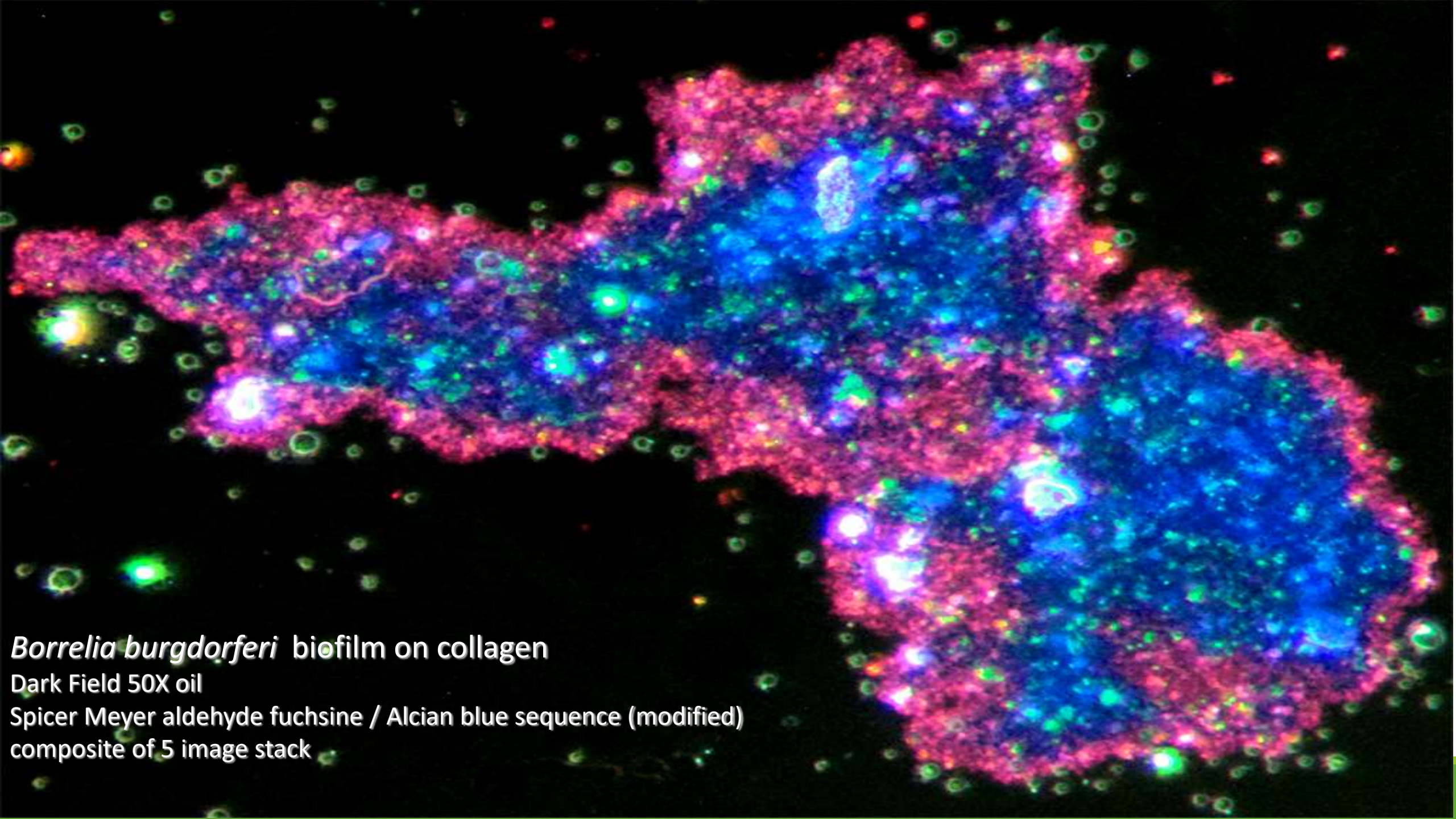
Stains acidic sulfated mucins

## Alcian blue:

Stains remaining non-sulfated mucins, carboxylated mucopolysaccharides

Spicer & Meyer, 1960





*Borrelia burgdorferi* biofilm on collagen  
Dark Field 50X oil  
Spicer Meyer aldehyde fuchsine / Alcian blue sequence (modified)  
composite of 5 image stack

# *Borrelia* EPS component: alginate?

---

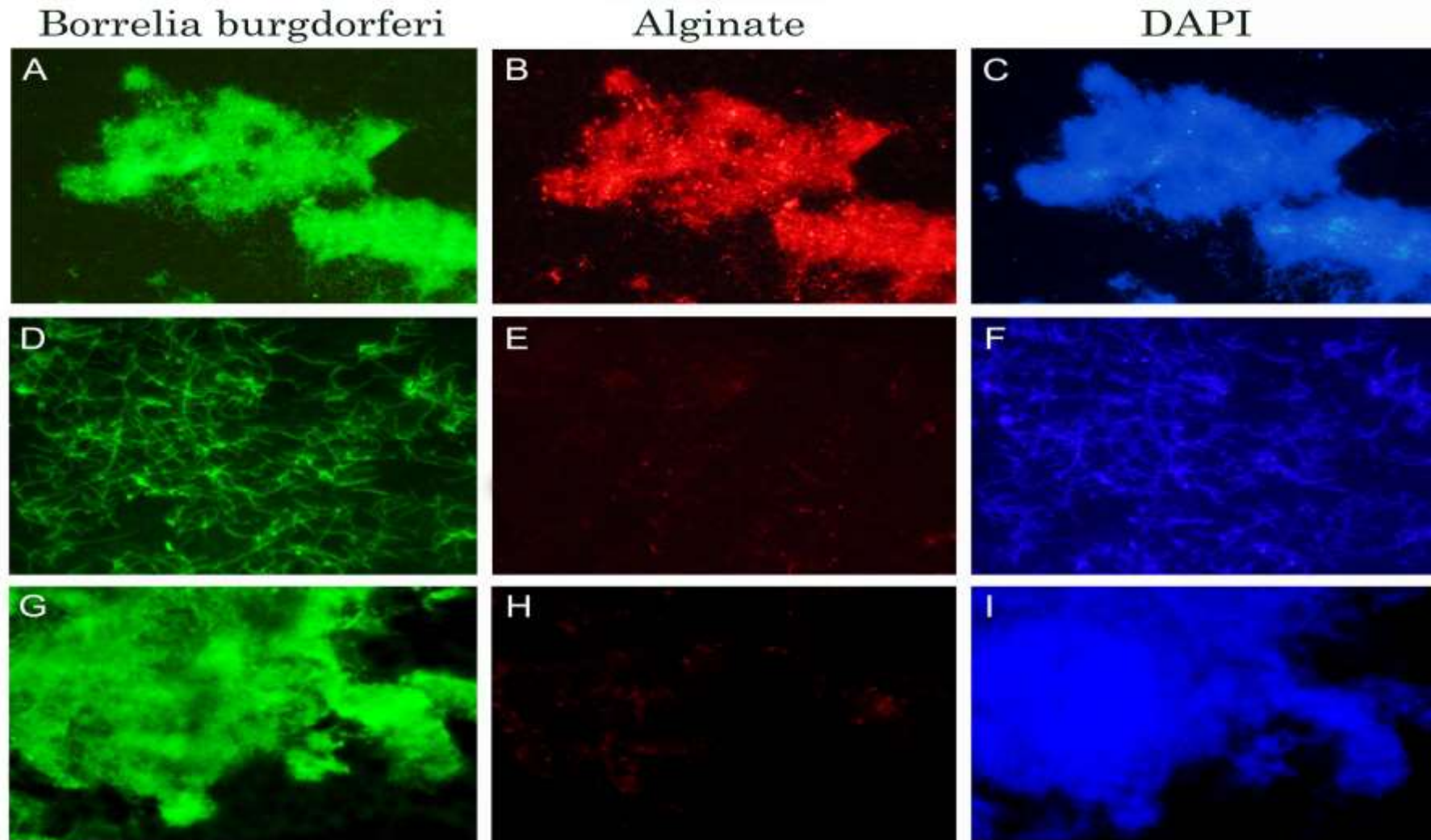
**Alginate** is well established as a viable primary EPS compound

Composed primarily of **polyuronic acid polymer – alginate** (Kjelleberg S 2007)

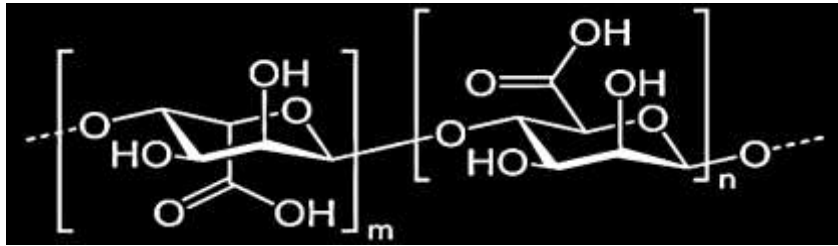
Chemical analysis of two varieties of slime produced by *Pseudomonas aeruginosa*  
(Murakama K 1973)

“The exopolysaccharide alginate protects *Pseudomonas aeruginosa* biofilm bacteria from macrophage killing.” (Leid JG 2005)

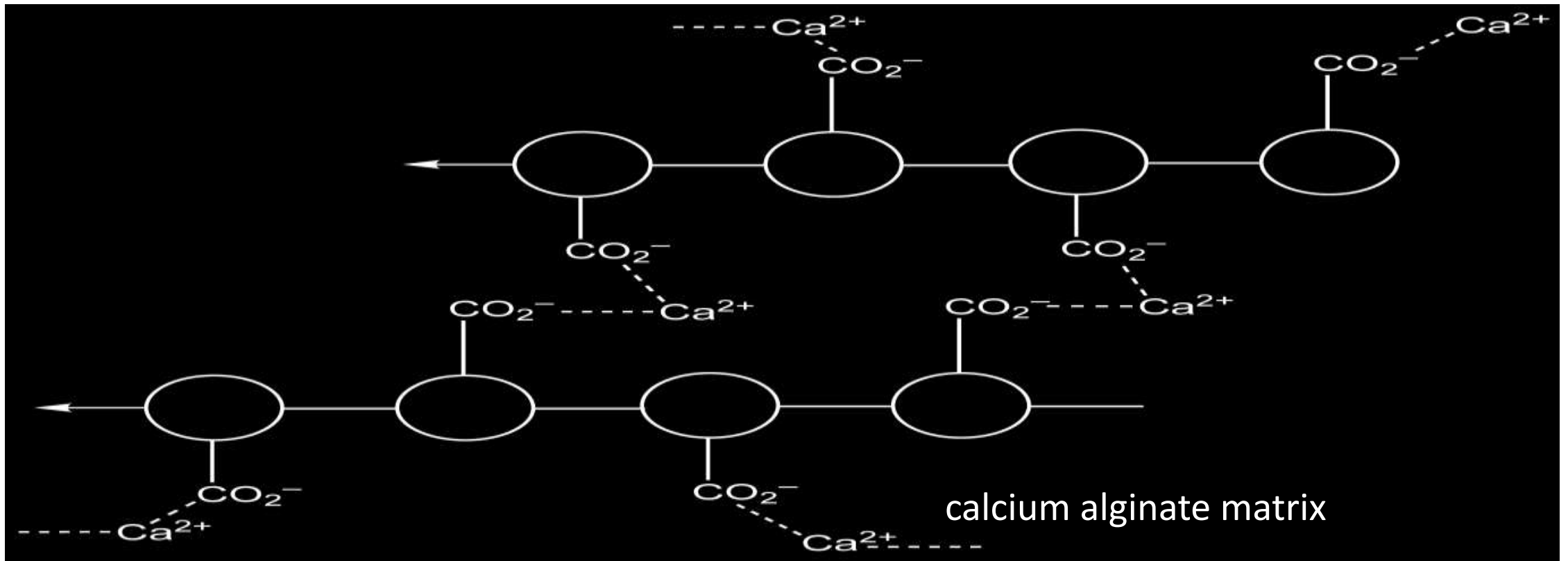
# Alginate on the surfaces of *Borrelia* aggregates



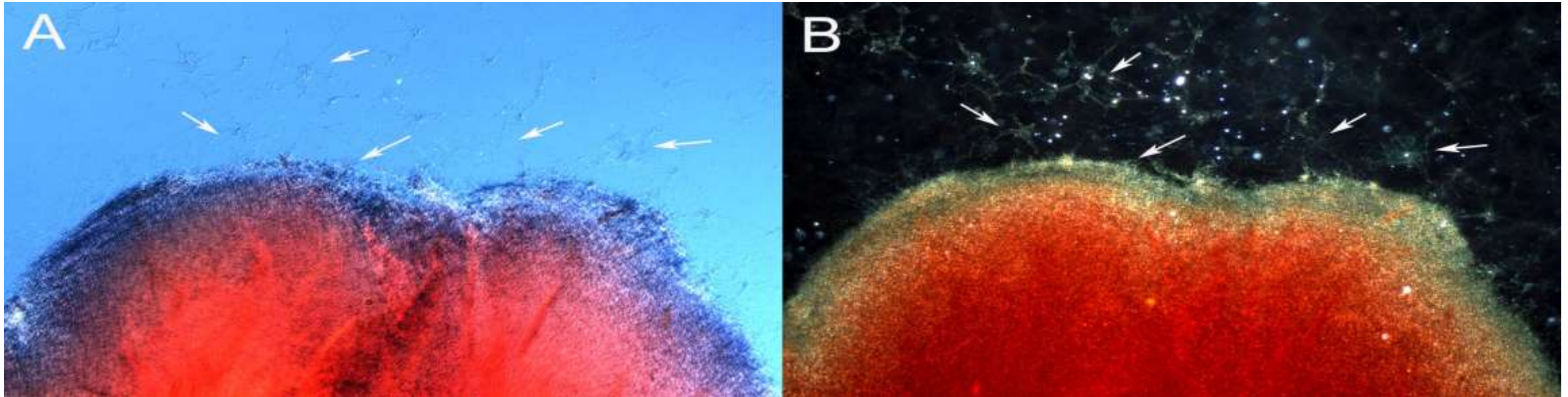
# EPS: alginate and calcium



copolymer subunits

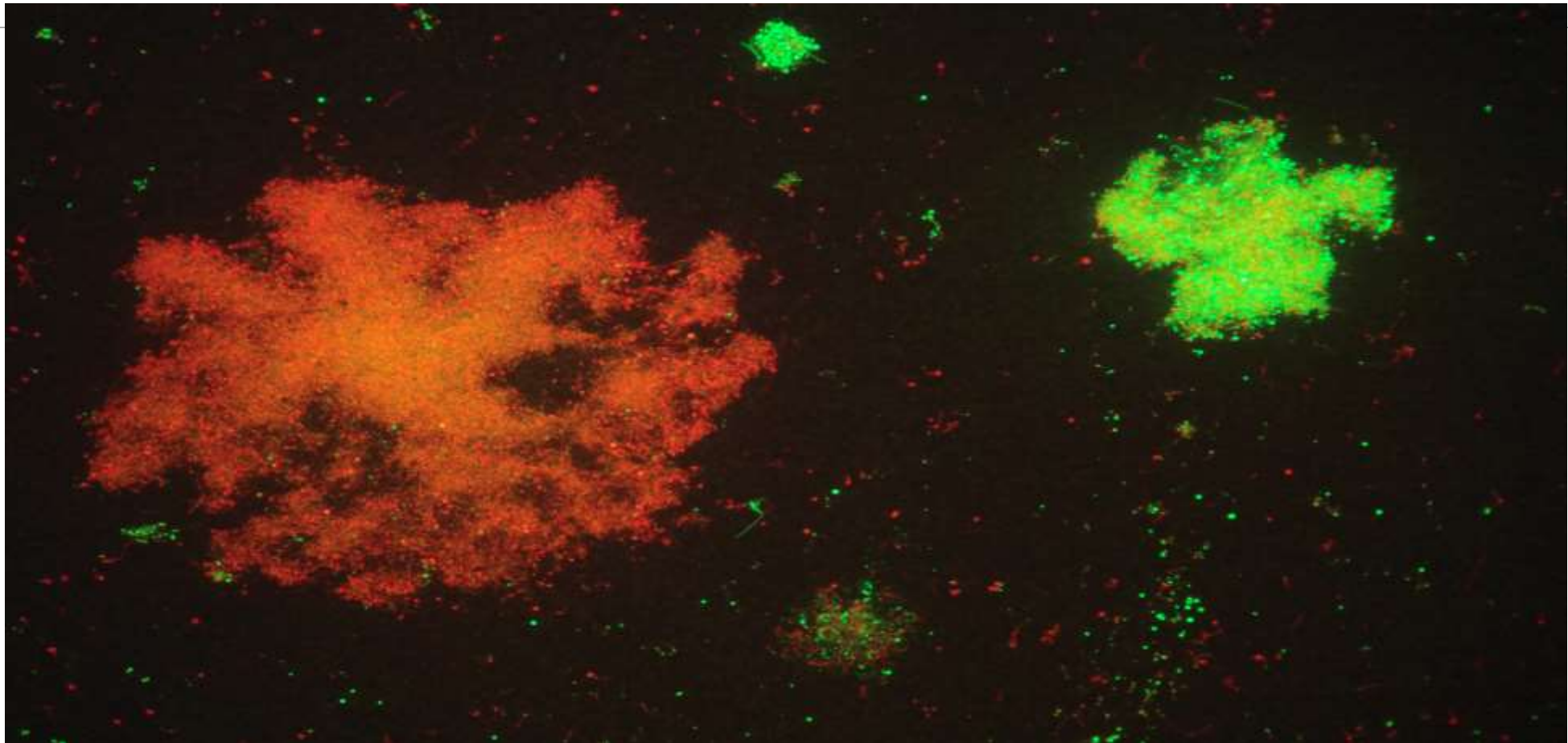


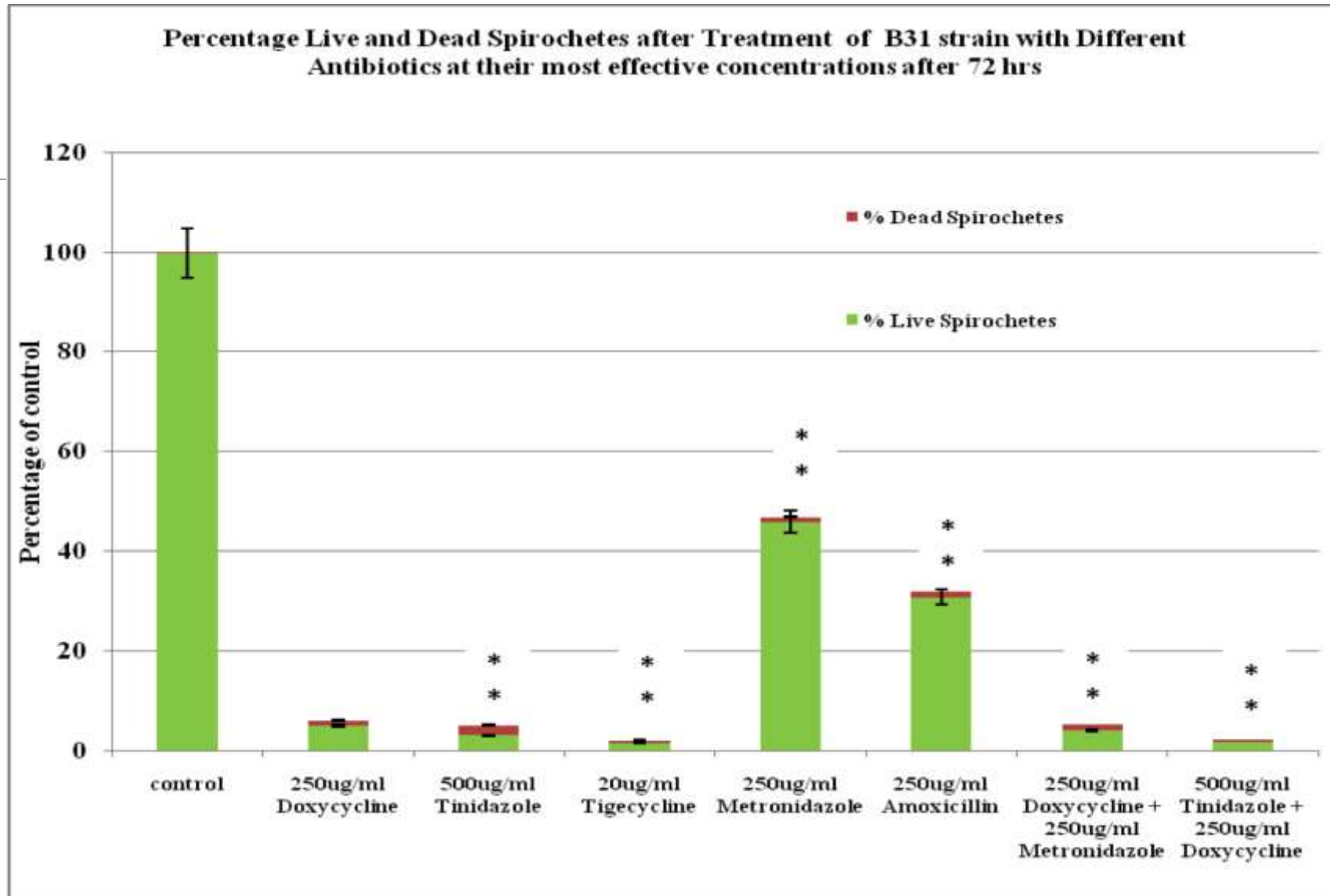
# Calcium on the surface of *Borrelia* aggregates



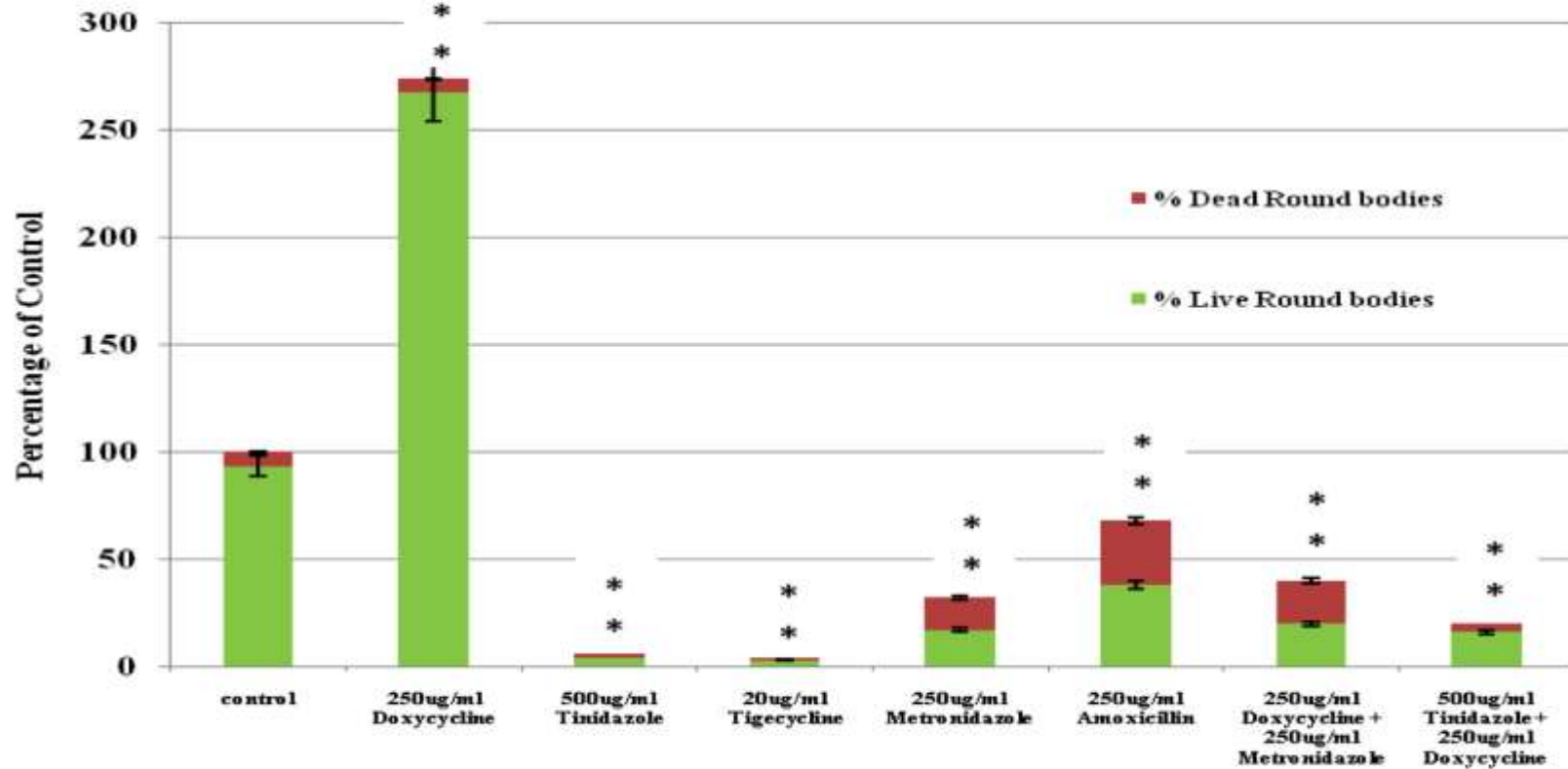
*Borrelia burgdorferi* B31 strain large aggregate surrounded by individual spirochetes and several small aggregates stained with the calcium-detecting stain Alizarin. Red coloration indicates presence of calcium, by differential interference contrast (Panel A) and dark field microscopy (Panel B).

*Borrelia burgdorferi* treated with 25 microgram/ml of doxycycline for 3 weeks



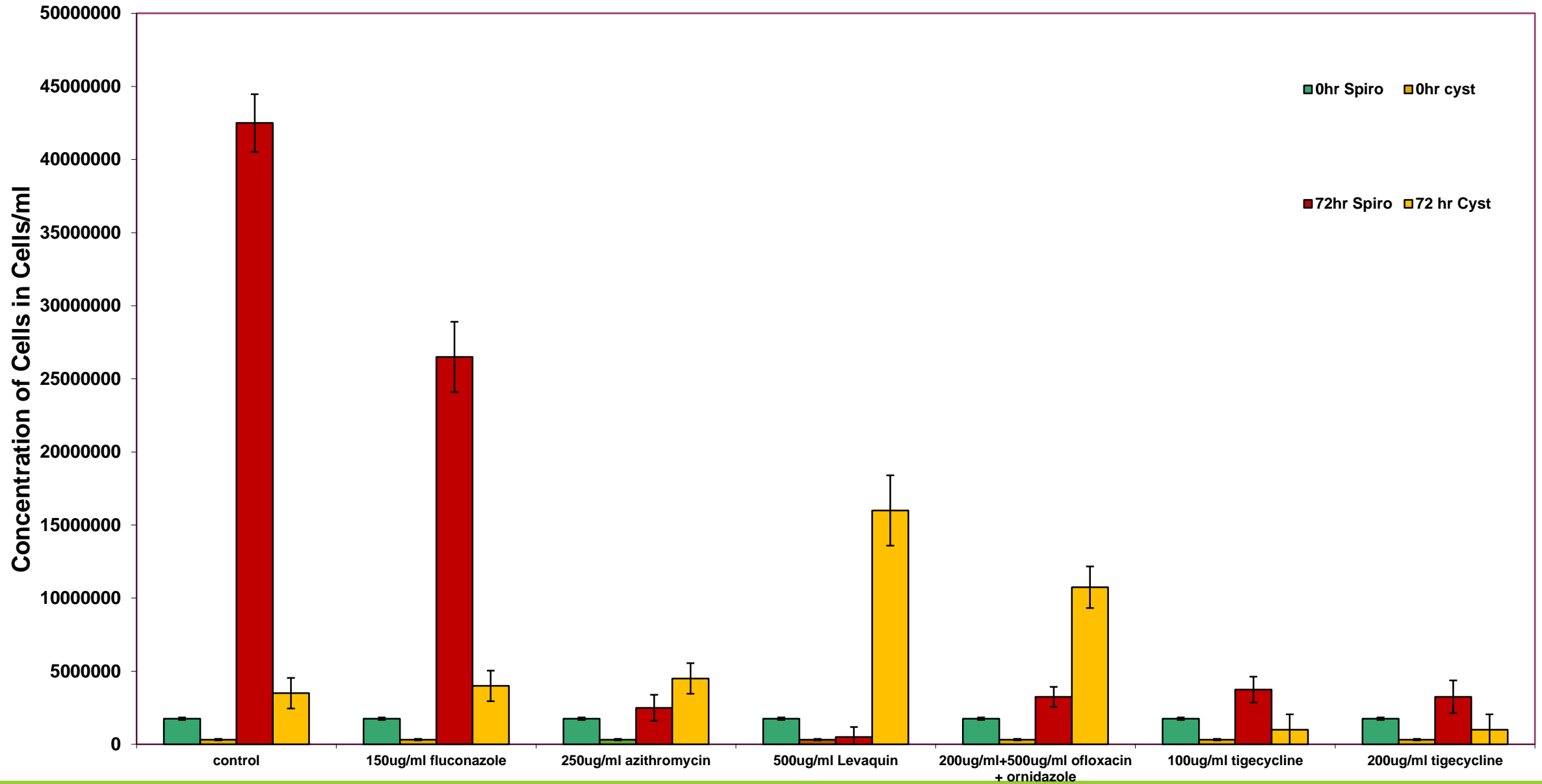


Percentage Live and Dead Round bodies after Treatment of B31 strain with Different Antibiotics at their most effective concentrations after 72 hrs

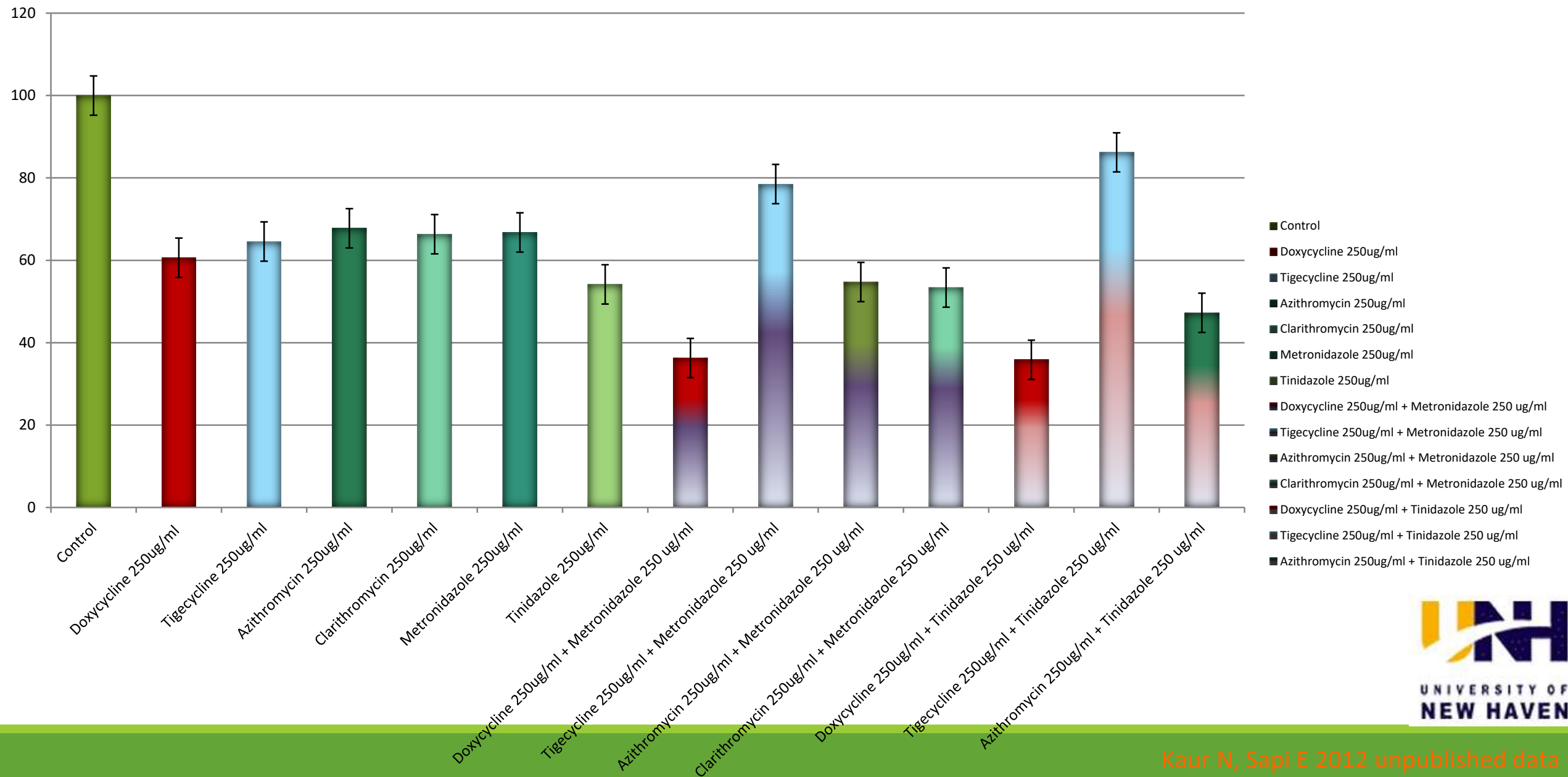


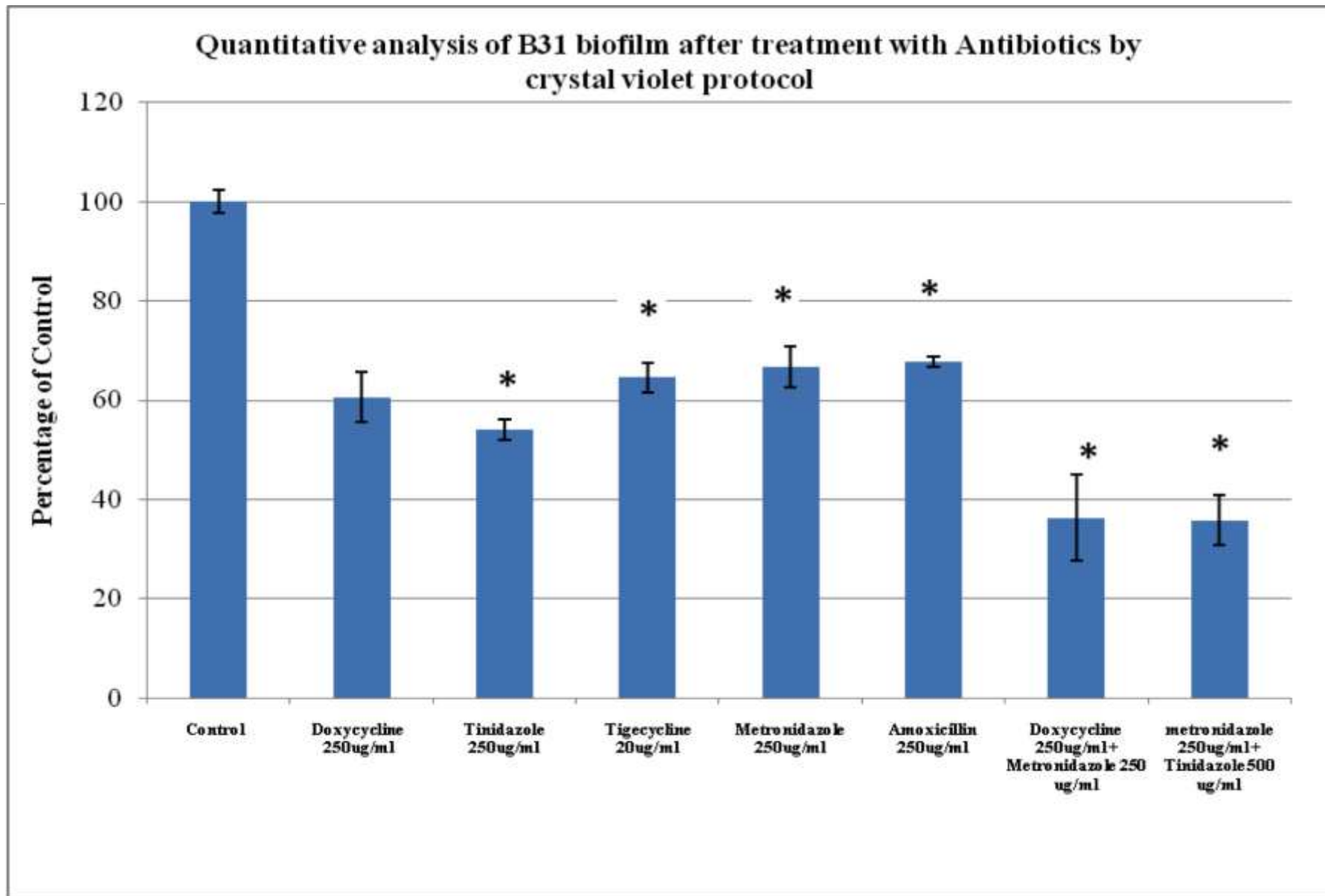


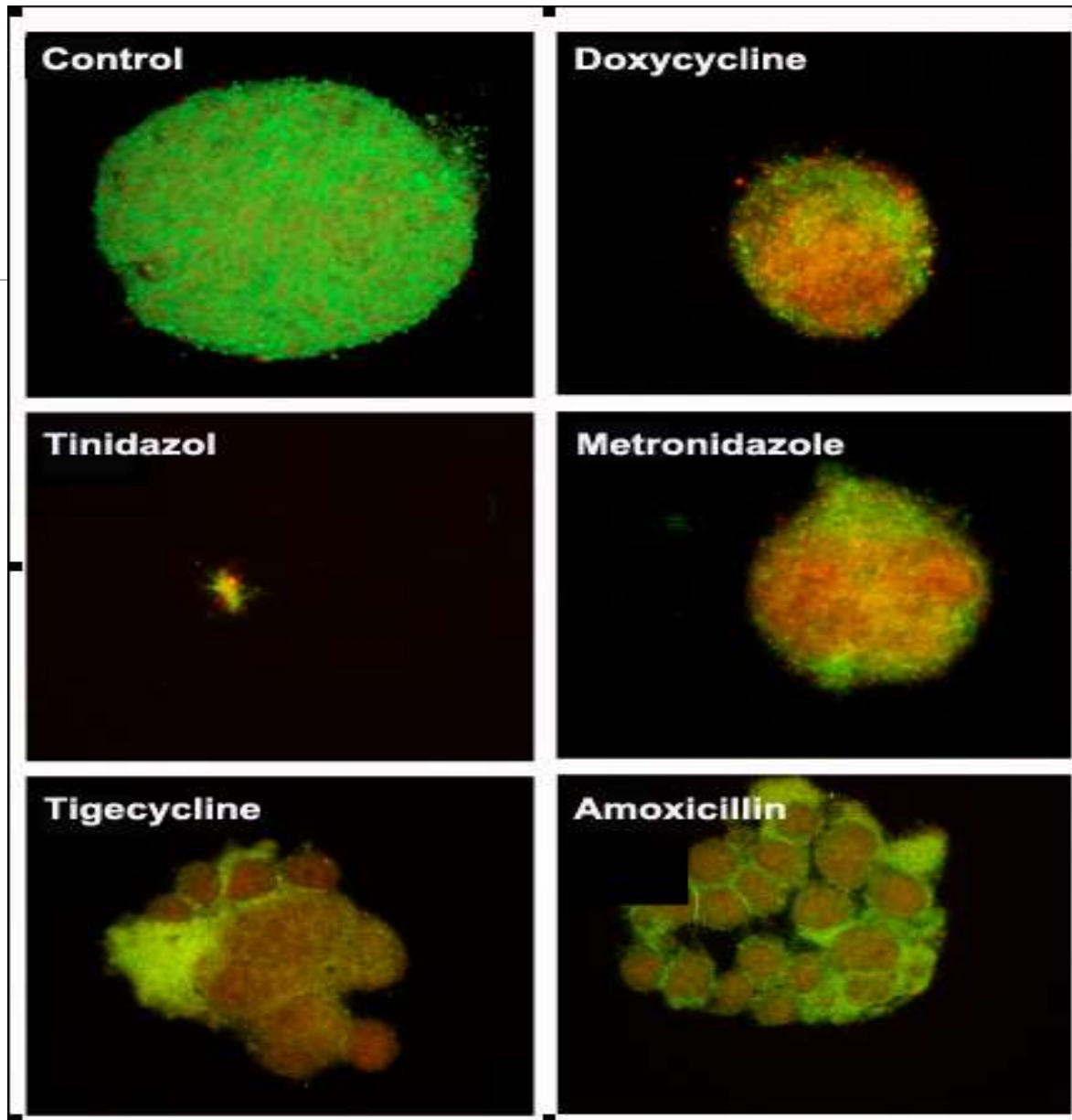
# Sensitivity of Borrelia forms to different antibiotics



# Effect of different antibiotics after treatment of Bb biofilm for 3 days





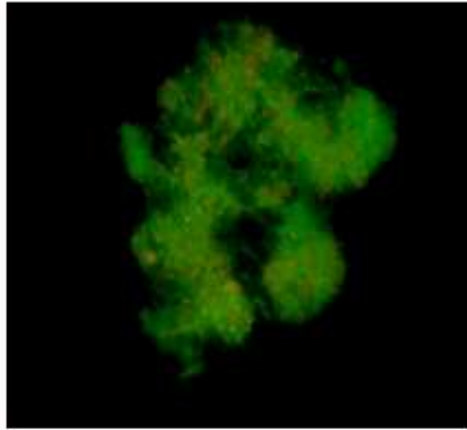


Red stain: Dead  
Green stain: Viable

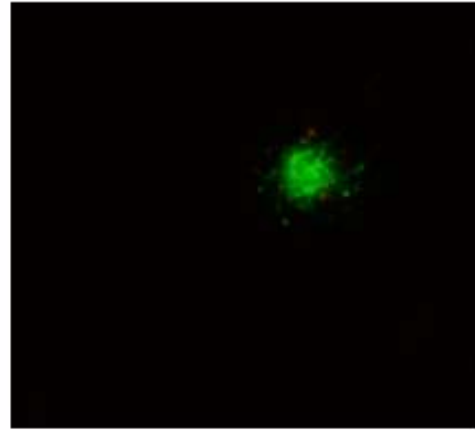
# Combination treatment

---

CONTROL



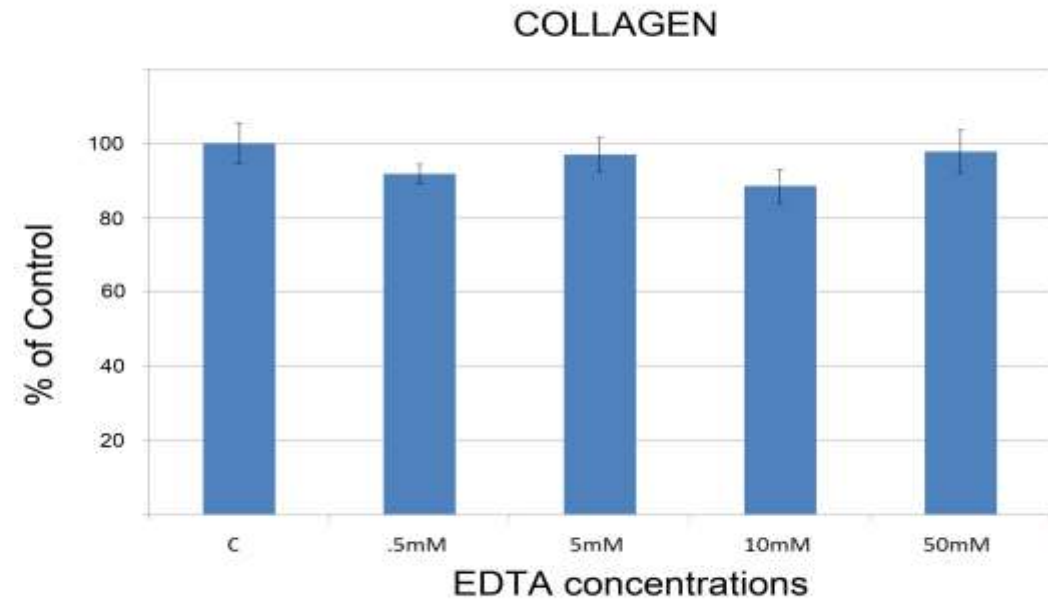
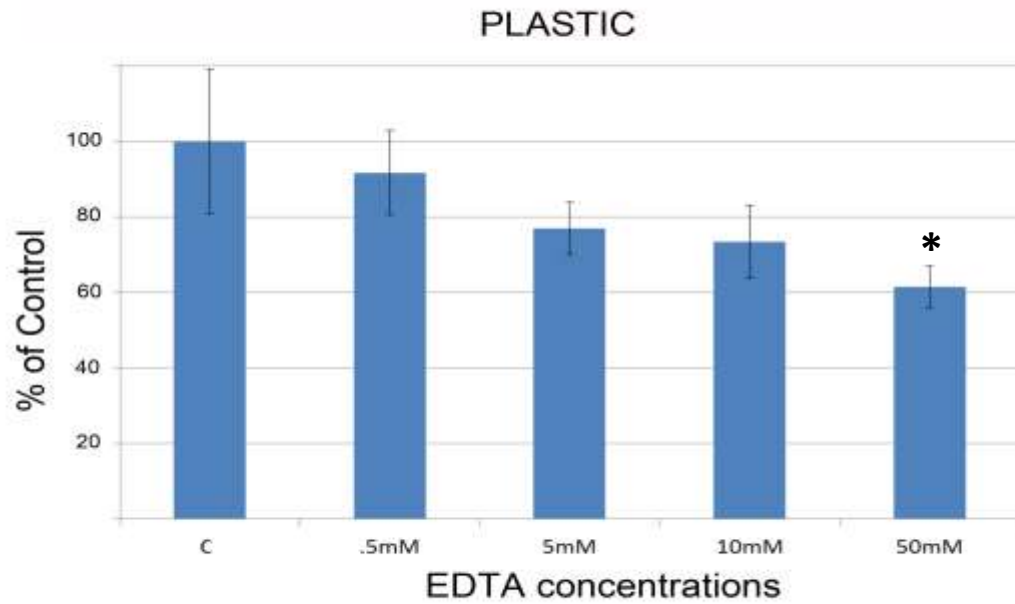
DOXY+  
METRONIDAZOLE



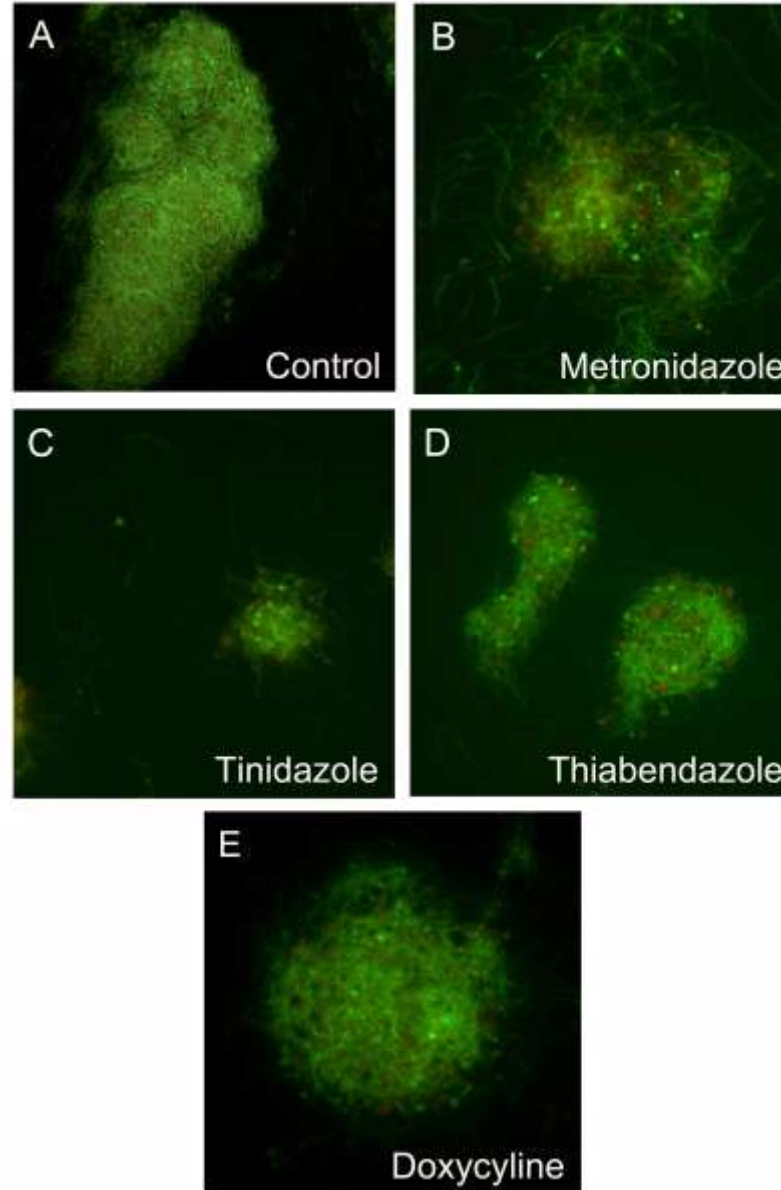
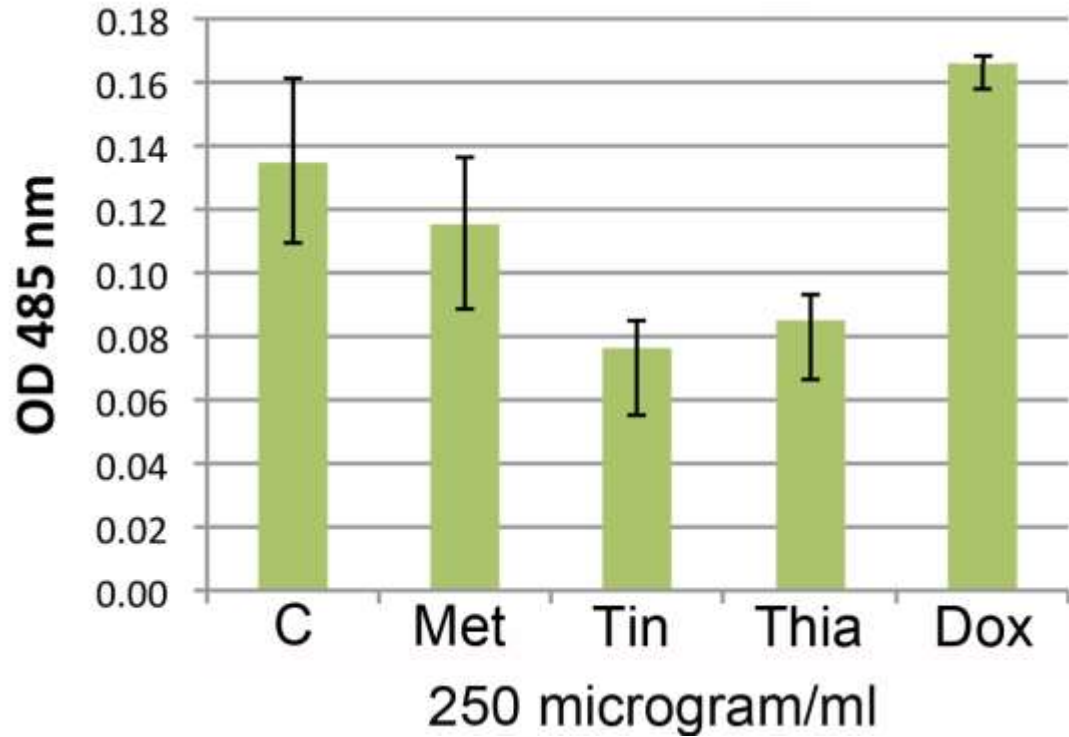
DOXY+  
TINIDAZOLE



# EDTA effect on *Borrelia* biofilm on plastic and on collagen - 48h



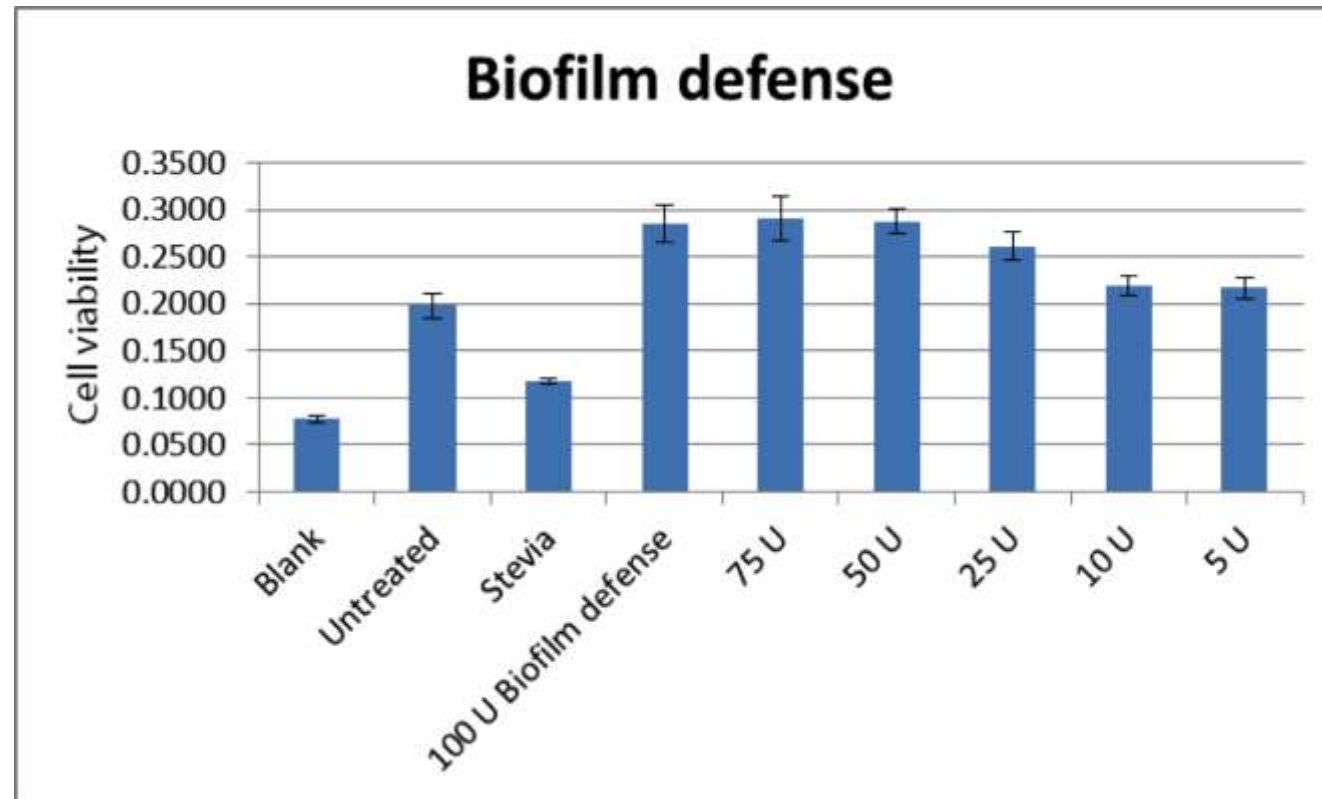
# Effects of different *nitroimidazole* antibiotics on *Borrelia* biofilm (72h)



# Mixture of Enzymes?

## Biofilm Defense

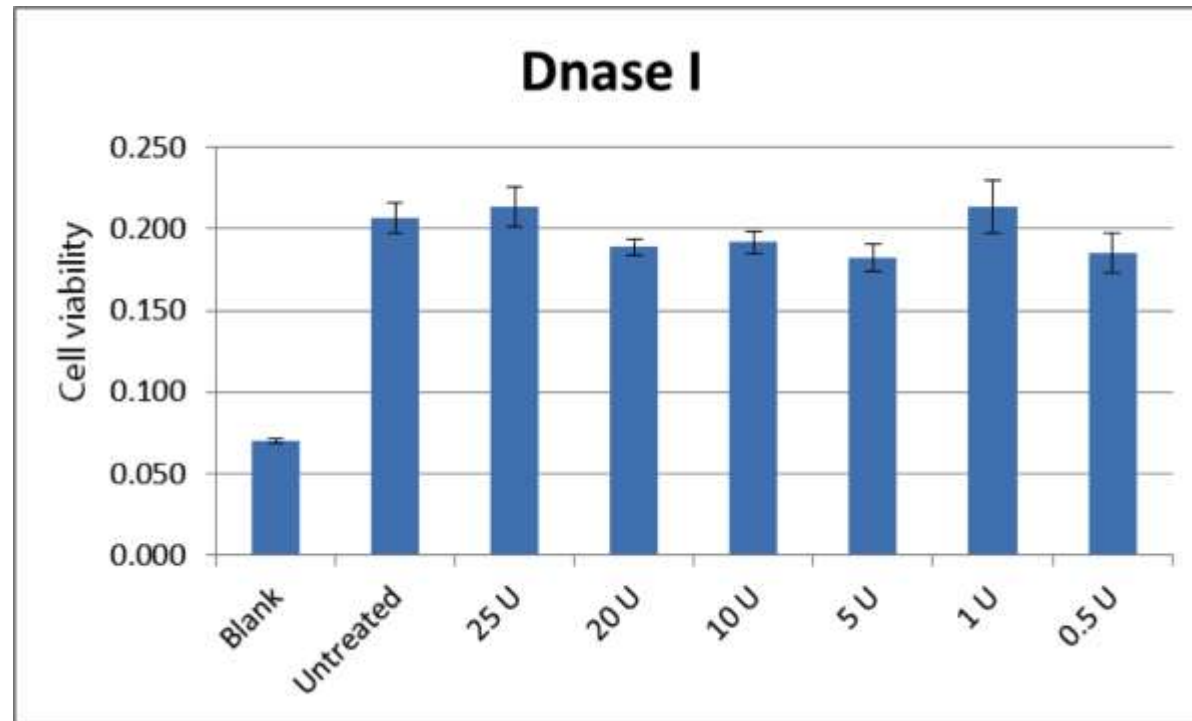
- NEC proprietary enzyme blend
- Pectinase
- Glucanase
- Serrazimes (endo/exo protease)
- Nanozymes (endo/exo protease)
- Beta-glucanase
- Protease AM
- Lipase
- Protease fl
- Alpha-galactosidase
- Xylanase
- Cellulase
- Amylase
- Invertase





# Degrading the extracellular DNA layer?

---

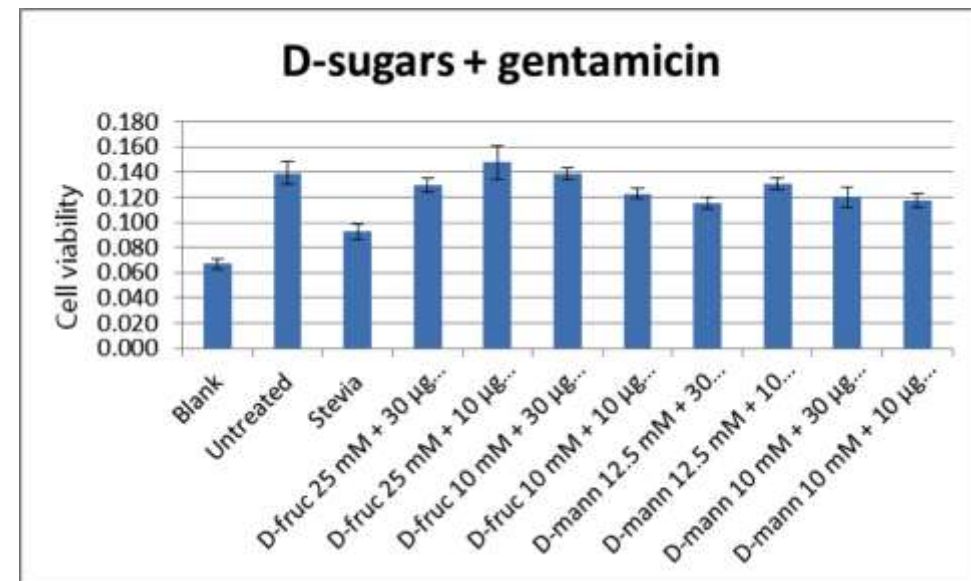
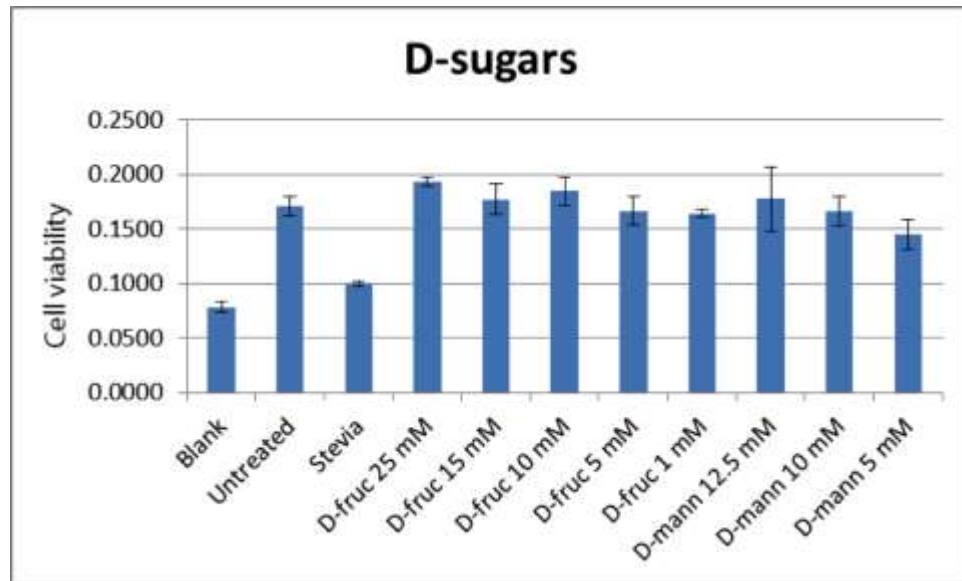


# Do we need a Trojan Horse?

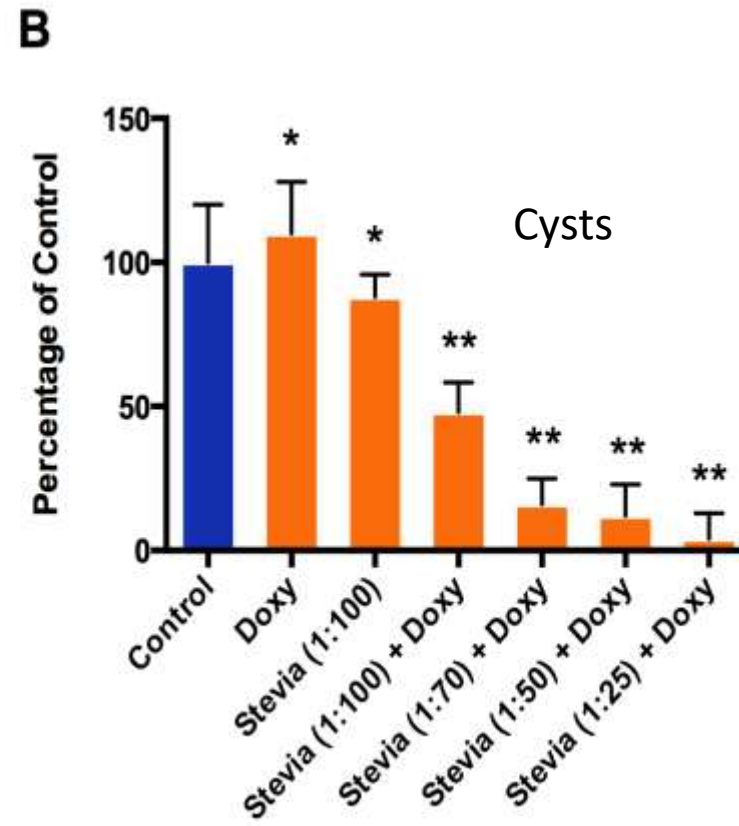
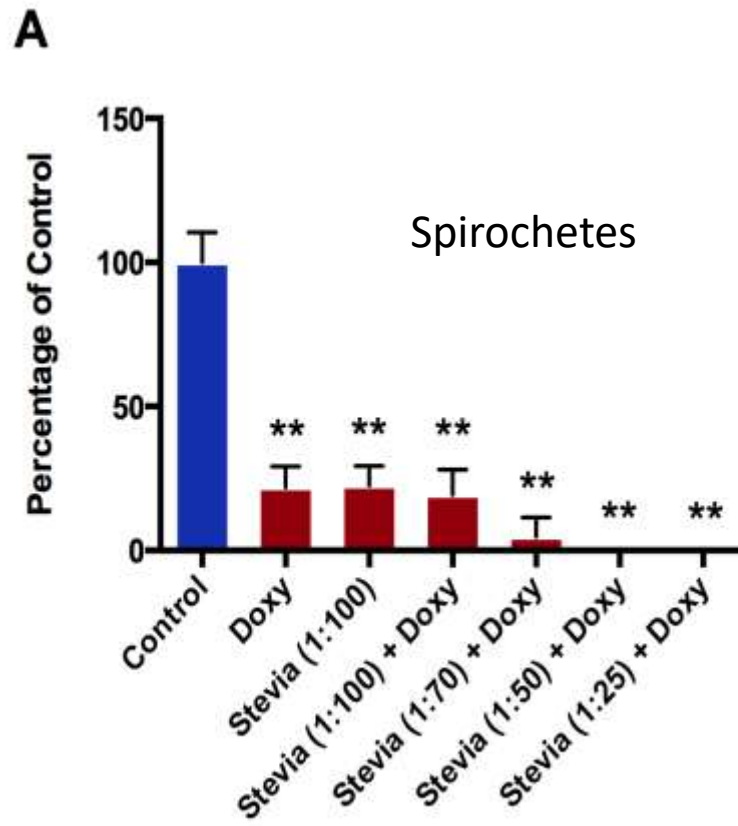
---



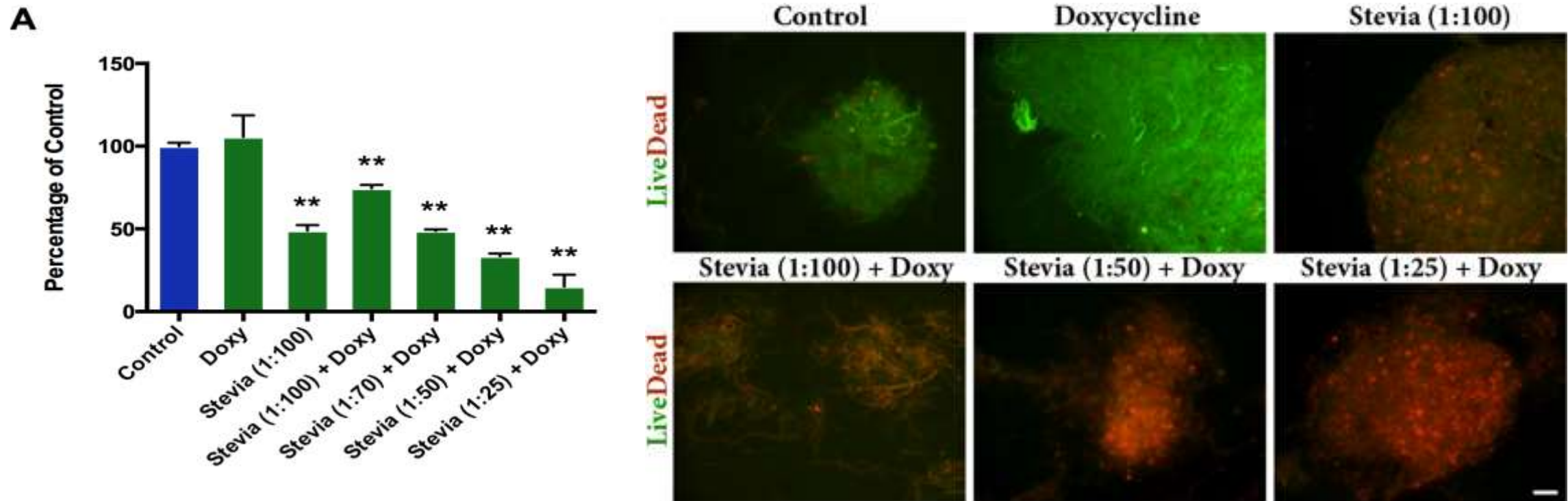
# Going after the persisters with sugar and aminoglycosides (Allison KR et al Nature 2011)



# Combining Stevia and Doxycycline?



# How about biofilm form?



# What is the next step?

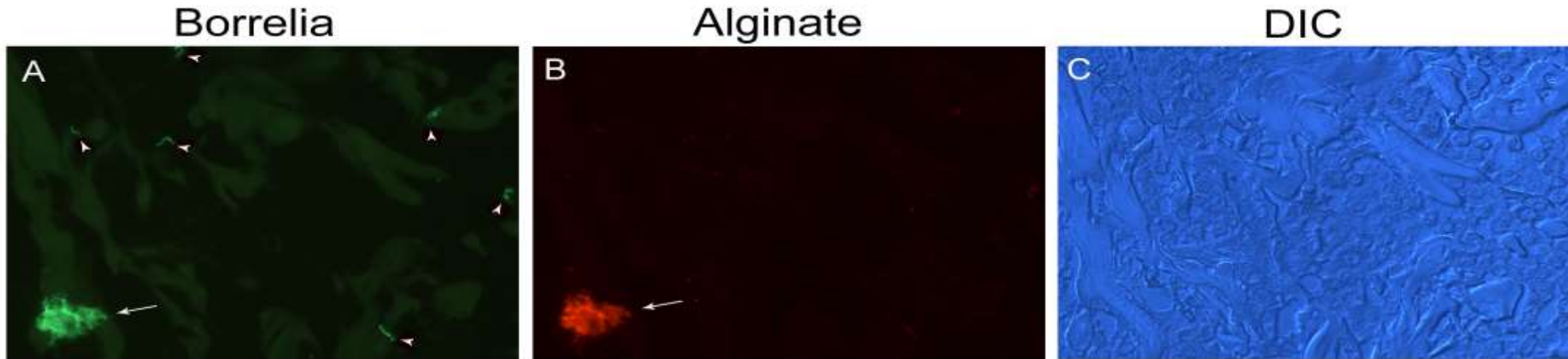
---

To demonstrate that *Borrelia* biofilms exist *in vivo*

- Skin biopsies from Borrelial lymphocytoma lesions (*Dr B Zelger*)

# Double immunostaining: **Borrelia** and **Alginate** antibody – Skin biopsies

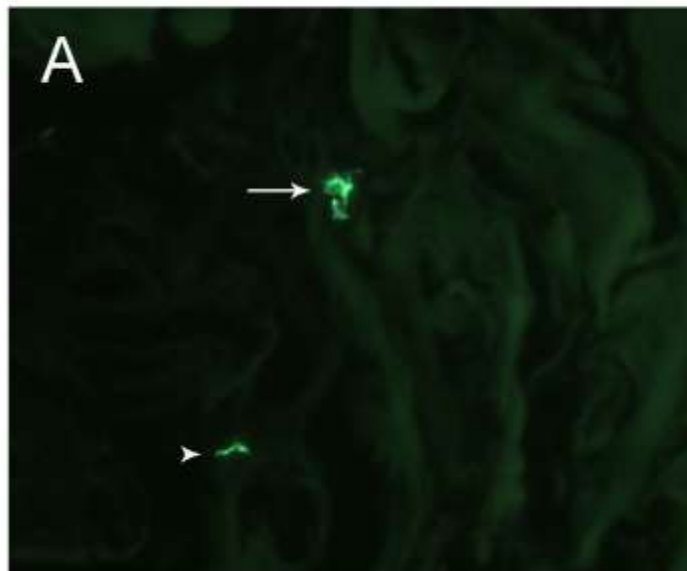
---



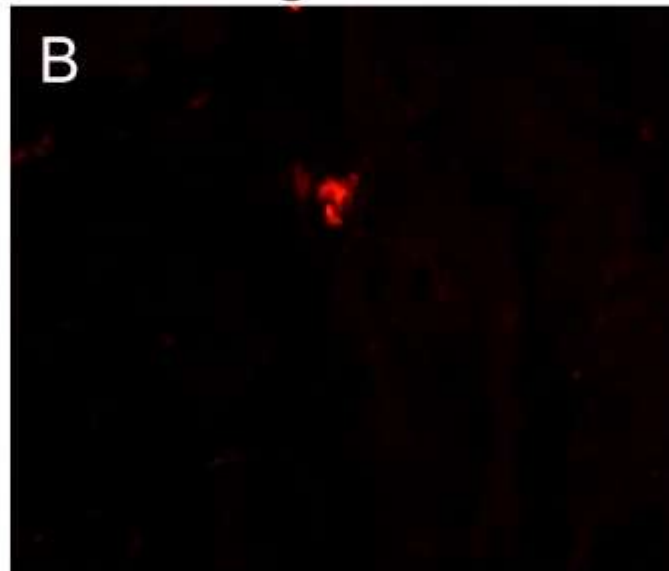
# Additional findings

---

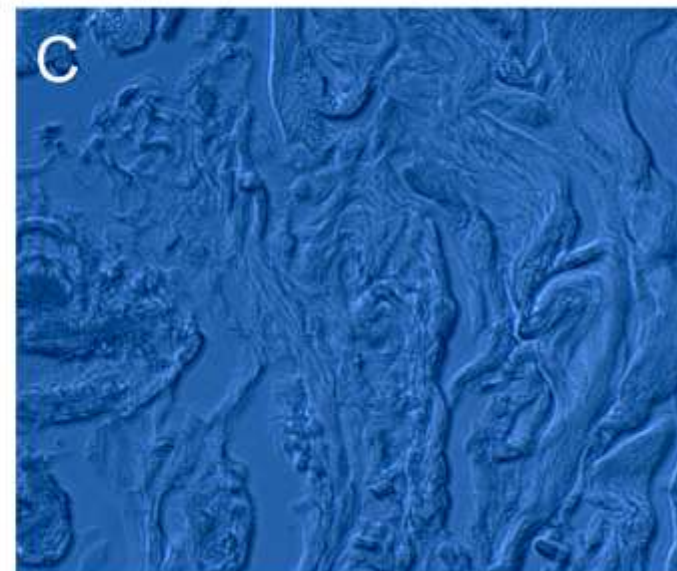
Borrelia



Alginate



DIC



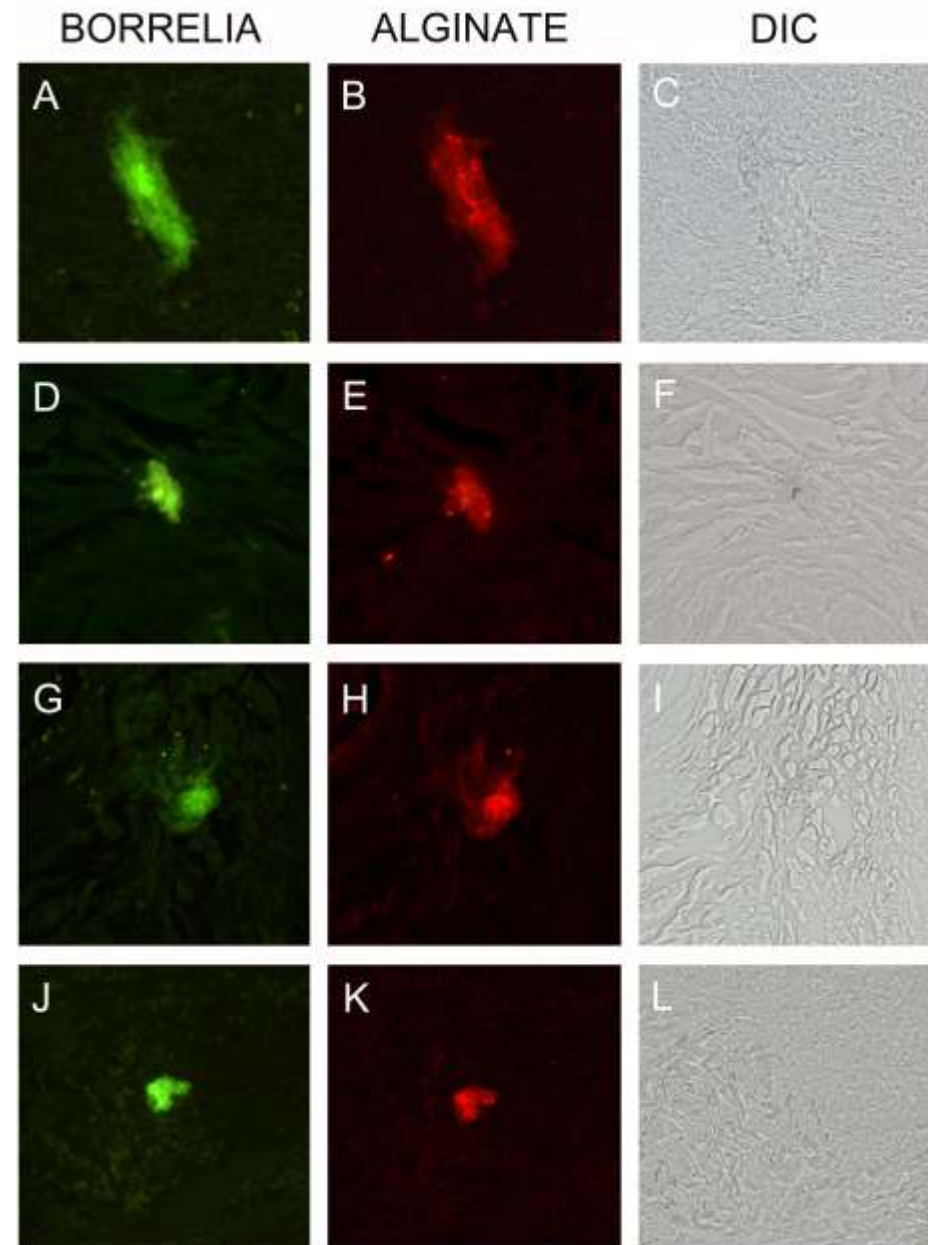


# Immunohistochemical images of infected skin samples

Borrelia antigen (green staining) and alginate (red).

DIC= differential Interphase differential microscopy

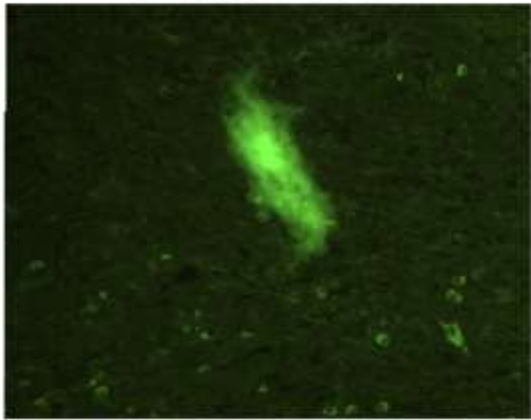
All samples were confirmed with PCR/direct sequencing for Borrelia



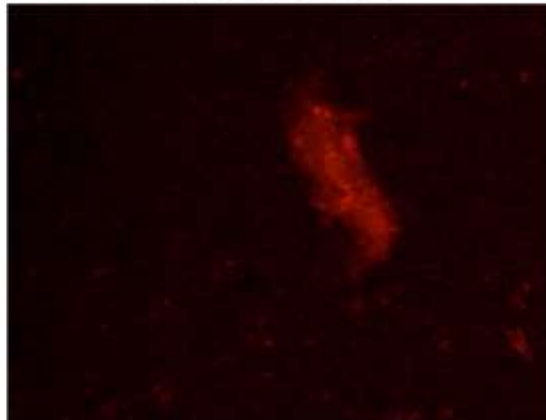
# *Borrelia* biofilm with silver staining proof in infected skin tissue

---

BORRELIA



ALGINATE



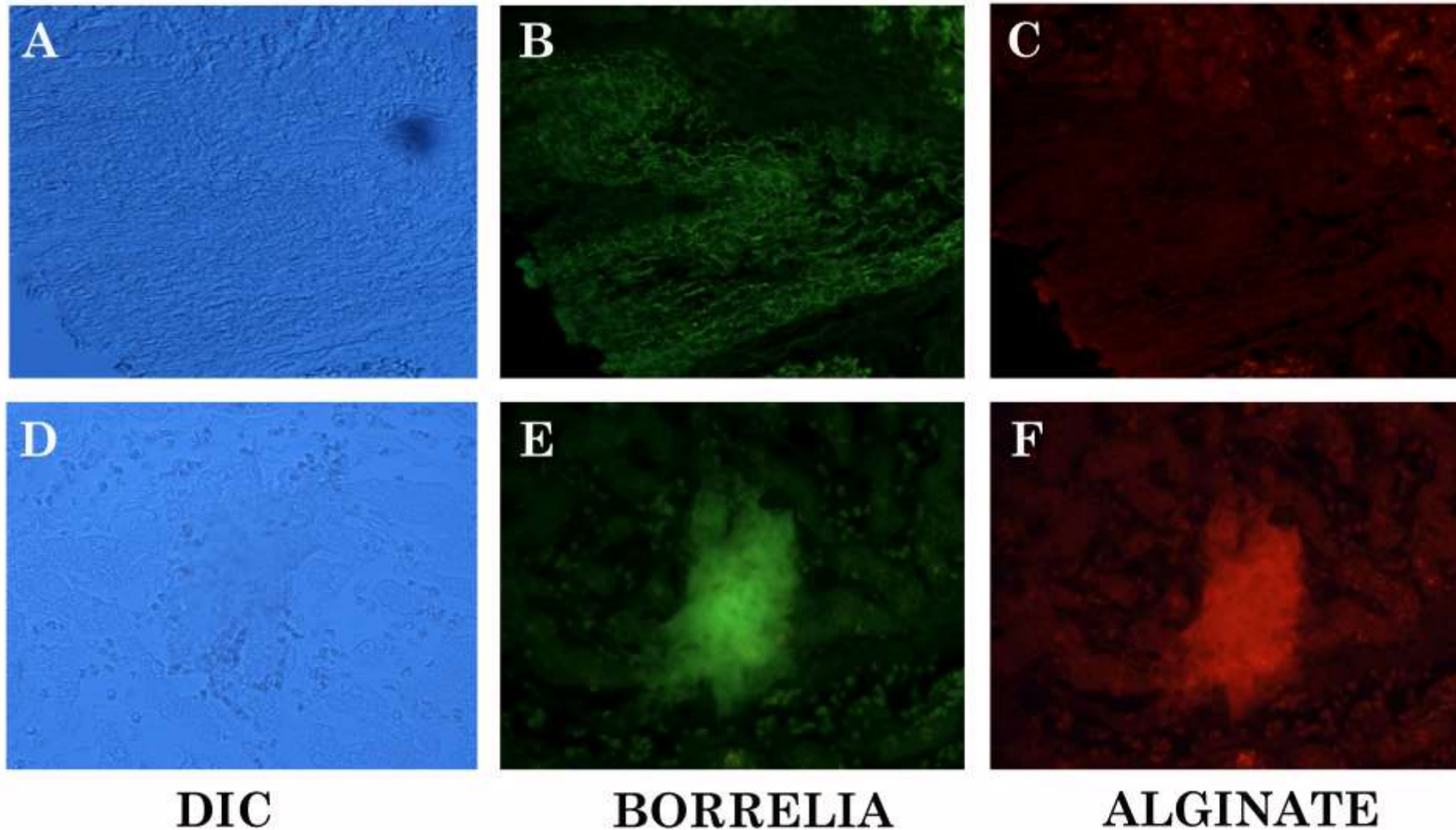
DIC



SILVER

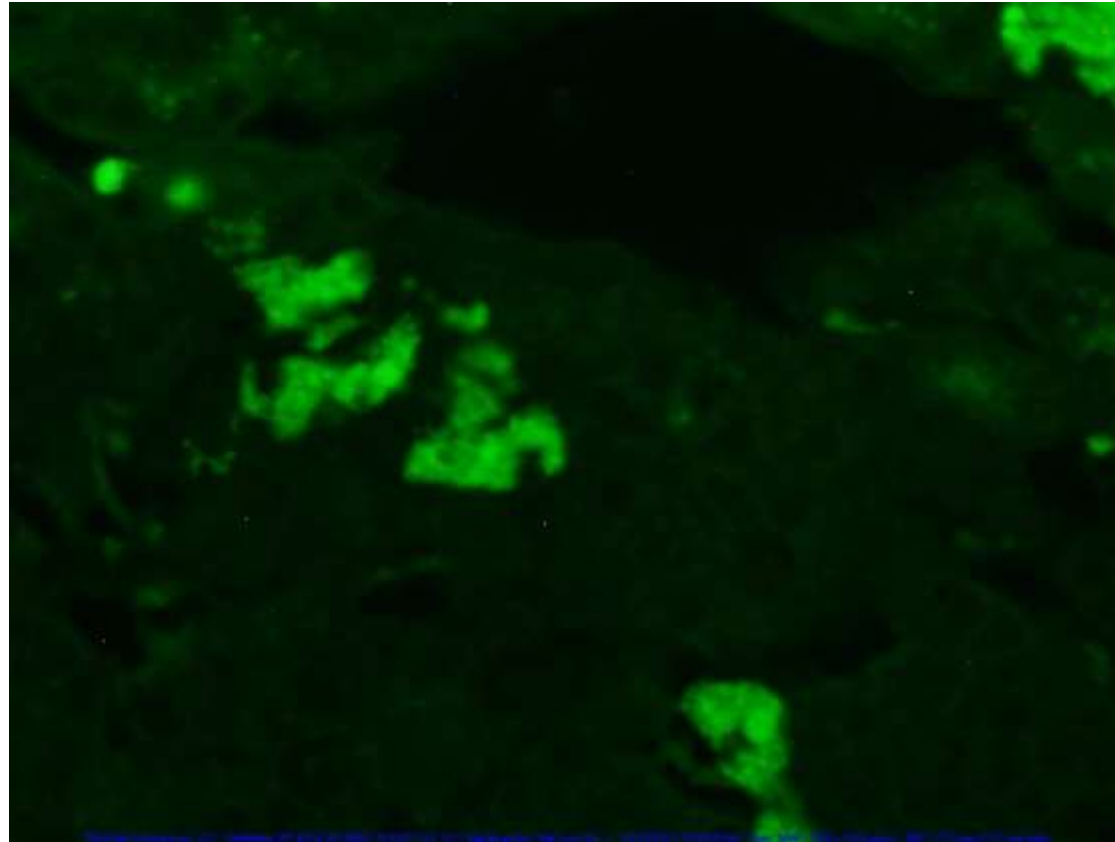


Where else can we find *Borrelia* biofilm in the body?



# Bb Infected Liver (MacDonald A, Middleveen M, 2014)

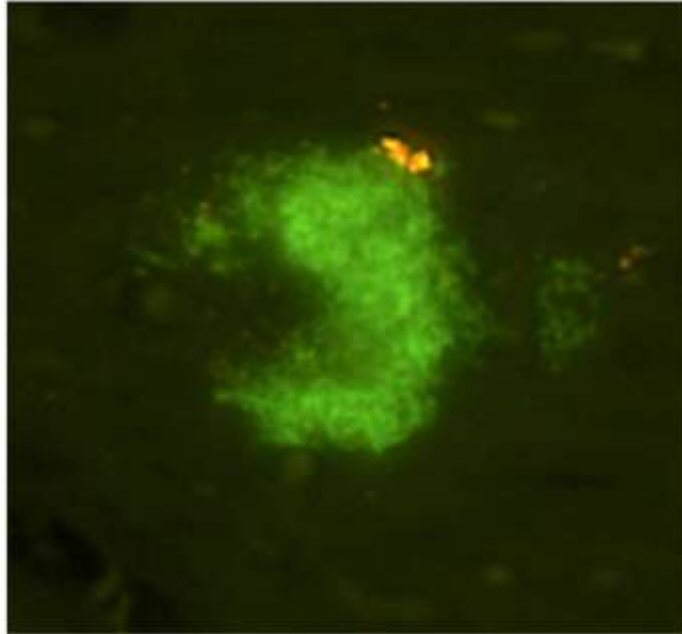
---



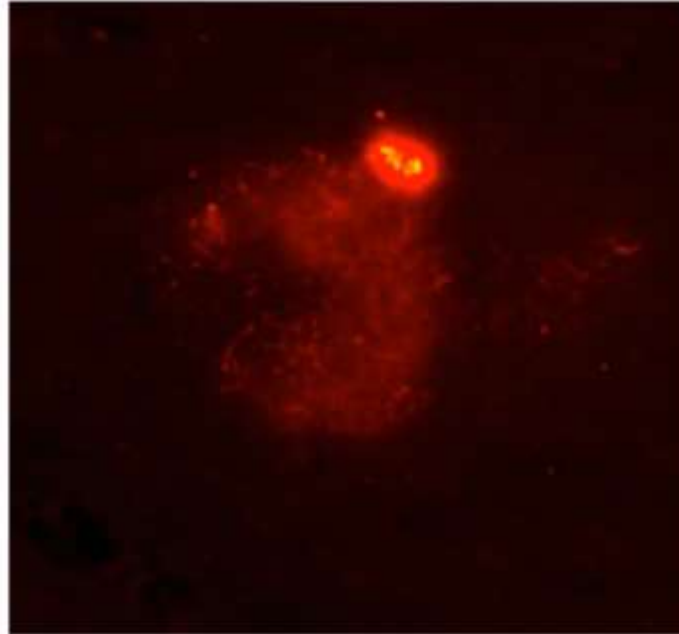
# Mouse Heart Tissue infected by Borrelia

---

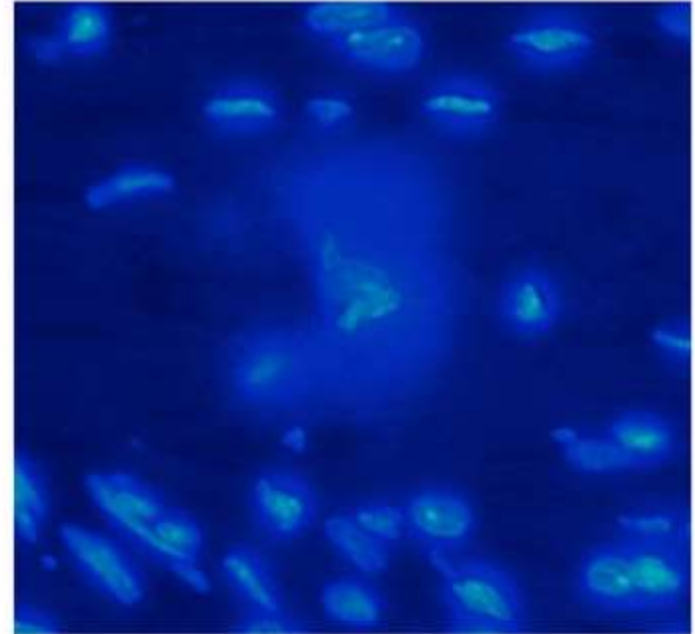
BORRELIA



ALGINATE



DAPI



# Summary

---

*Borrelia burgdorferi* does form a very organized biofilm *in vitro*

Major component is probably calcium alginate, but polymer subunits remain to be identified

Biofilm provide a refuge from antimicrobial treatment

Preliminary results show potential *in vivo* *Borrelia* biofilm in infected skin tissues as well other tissues

# The new atomic force microscope UNH Lyme disease research group



# Special Thanks To:



---

University of New Haven and College of A&S for funding our studies

Lymedisease.org, Global Lyme Alliance, Lyme Disease Association, TBDA (Turn the Corner Foundation), Portman Foundation, Wartman Family for supporting our research projects

Lymedisease.org, Lyme Research Alliance, LDA, TBDA (Turn the Corner), and Schwartz Foundation for providing a “state of the art” microscopes for our morphological studies

Dr. Michael Rossi and Dr. Roman Zajac at the Department of Biology and Environmental Science (UNH) for additional support and funds

Dr. Alan MacDonald for his never-ending support



# Question?

---

Please call me or send me an email:

[esapi@newhaven.edu](mailto:esapi@newhaven.edu)

1-203-479-4552